



VIII. Sample Preparation:

• Serum

1. Add 30 μ l of Serum Solution into 270 μ l of serum in an Eppendorf tube and vortex well.
2. Incubate the sample at 37 °C for 45 min.
3. After 45 min, incubate the sample at 85-90 °C for 10 min.
4. Dilute the sample 10 fold using the Sample Diluent. For example, mix 20 μ l of serum with 180 μ l of Sample Diluent.
5. Use 50 μ l per well for the assay.

Note: Dilution factor: 10

• Urine

1. Centrifuge 0.5 ml of urine at 10,000 x g for 5 min and collect the supernatant.
2. Dilute the supernatant 10 fold using the Sample Diluent. (For example, mix 20 μ l of urine with 180 μ l of Sample Diluent.)
3. Use 50 μ l per well for the assay.

Note: Dilution factor: 10

• Milk

1. Add 20 μ l of Extraction Solution to 1 ml of milk and vortex well.
2. Centrifuge the sample at 10,000 x g for 20 min at 4 °C and collect the clear supernatant.
3. Dilute the supernatant 10 fold with Sample Diluent. For example, mix 20 μ l of the supernatant with 180 μ l of Sample Diluent.
4. Use 50 μ l per well for the assay.

Note: Dilution factor: 10

IX. Quinolone ELISA Assay Protocol:

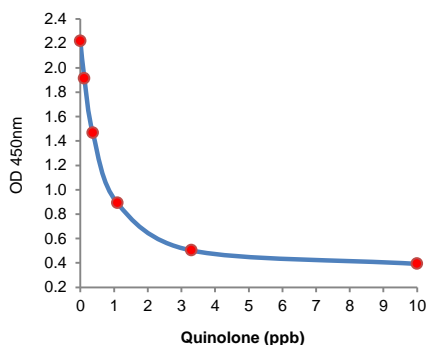
Notes: It is recommended that all standards and samples should be run at least in duplicate. Standard curves must be run each time an assay is performed.

1. Prepare all reagents, Standards and samples as sections VII and VIII respectively.
2. Add 50 μ l of Standards or Samples per well. Then add 50 μ l of HRP-conjugate and 50 μ l of Antibody to the above wells.
3. Cover the microtiter plate with plate sealer and mix well. Incubate the plate at room temperature (25°C) for 60 min.
4. Aspirate all reagents and wash each well 4 times. Add 250 μ l of 1X Wash Buffer and incubate for 30 seconds. Remove 1X Wash buffer completely before the next wash. **Note:** This is essential for accurate results. Repeat this step 3 more times.
5. Add 100 μ l of TMB Substrate to each well. Tap or shake the plate to ensure complete mixing.
6. Check the OD at 650 nm for the well containing no Quinolone (S0). When its reading is between 0.95 and 1.05 (usually between 10-30 min after adding the TMB Substrate), add 50 μ l of Stop Solution and gently tap the plate to ensure thorough mixing.
7. Measure the OD at 450 nm for the Standards and samples within 10 min.

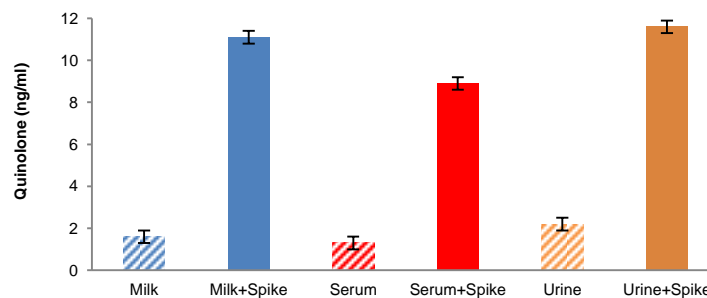
X. Calculation:

The Standard Curve is done by plotting the OD at 450 nm vs. Quinolone concentrations. The concentration of Quinolone in each sample (ng/ml), which can be read from the calibration curve, is multiplied by the corresponding dilution factor.

A.



B.



Figures. A. Quinolone Standard Curve for Quinolone ELISA Kit (*This standard curve is for demonstration only. A Standard Curve must be run with each assay*). **B.** Spike recovery experiment: Human serum, urine and milk samples were assayed without and with spike of 10 ng/ml quinolone. Recovery rate: 80-100%.

XI. Related Products:

Gentamicin (serum/urine) ELISA Kit (Cat. No. K4315-100)
Ampicillin ELISA Kit (Cat. No. E4350-100)
Enrofloxacin (ENR) ELISA Kit (Cat. No. E4277-100)

Folic Acid ELISA Kit (Cat. No. 4523-100)
Kanamycin ELISA Kit (Cat. No. K4210-100)

FOR RESEARCH USE ONLY! Not to be used on humans.