



# Vanillylmandelic Acid ELISA Kit

(Catalog# E4752-100; 96 assays; Storage at 4°C)

## I. Introduction:

Vanillylmandelic acid (VMA), a metabolic by-product of norepinephrine and epinephrine, can be used to detect neuroblastoma and other tumors of neural crest origin. BioVision's Vanillylmandelic acid ELISA kit is based on the Competitive-ELISA principle. The micro ELISA plate provided in this kit has been pre-coated with VMA. During the reaction, VMA in the sample or standard competes with a fixed amount of VMA on the solid phase supporter for sites on the Biotinylated Detection Ab specific to VMA. Excess conjugate and unbound sample or standard are washed from the plate, and Avidin conjugated to Horseradish Peroxidase (HRP) are added to each microplate well and incubated. Then a TMB substrate solution is added to each well. The enzyme-substrate reaction is terminated by the addition of stop solution and the color change is measured spectrophotometrically at a wavelength of 450 nm ± 2 nm. The concentration of VMA in the samples is then determined by comparing the OD of the samples to the standard curve.

## II. Applications:

- Precision: Coefficient of variation is < 10%.
- Quantitative measurement of Vanillylmandelic Acid in serum, plasma and other biological fluids.
- Detection Range-1.56—100 ng/ml
- Specificity: This kit recognizes VMA in samples. No Significant cross-reactivity or interference between VMA and analogues was observed.
- Sensitivity: 0.94 ng/ml

## III. Sample Type:

- Plasma
- Serum
- Other biological fluids

## IV. Kit Contents:

Components	E4752-100	Part Number	Storage Temp
Micro ELISA Plate	8 wells x12 strips	E4752-100-1	-20°C
Standard	2 vials	E4752-100-2	-20°C
Biotinylated Detection Ab (100x)	120 µl	E4752-100-3	-20°C
HRP Conjugate (10x)	120 µl	E4752-100-4	-20°C (In dark)
Standard & Sample Diluent	20 ml	E4752-100-5	4°C
Biotinylated Detection Antibody Diluent	14 ml	E4752-100-6	4°C
HRP Conjugate Diluent	14 ml	E4752-100-7	4°C
Wash Buffer (25x)	30 ml	E4752-100-8	4°C
Substrate Reagent	10 ml	E4752-100-9	4°C (In dark)
Stop Solution	10 ml	E4752-100-10	4°C
Plate Sealer	4	E4752-100-11	RT

## V. User Supplied Reagents and Equipment:

- Deionized or distilled water
- Microplate reader with 450 nm wavelength filter

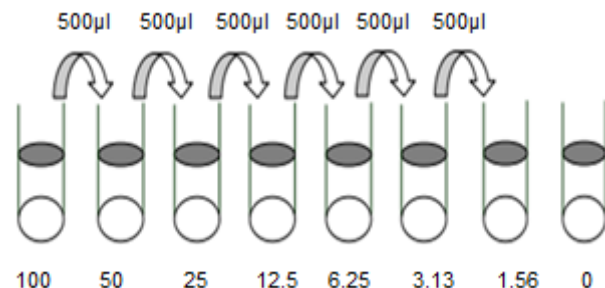
## VI. An unopened kit can be stored at 4°C for 1 month. If the kit is not used within 1 month, store the items separately according to the following conditions once the kit is received.

**Biotinylated Detection Antibody:** Calculate the required amount before the experiment (50 µl/well). Centrifuge the stock tube before use, dilute the 100x Concentrated Biotinylated Detection Ab to 1x working solution with Biotinylated Detection Ab Diluent Wash Buffer.

**HRP Conjugate:** Calculate the required amount before the experiment (100µl/well). Dilute the 100x Concentrated HRP Conjugate to 1x working solution with Concentrated HRP Conjugate Diluent.

**Wash Buffer:** Dilute 30 mL of Concentrated Wash Buffer with 720 mL of deionized or distilled water to prepare 750 mL of Wash Buffer. Note: if crystals have formed in the concentrate, warm it in a 40°C water bath and mix it gently until the crystals have completely dissolved.

**Standard Preparation:** Centrifuge the standard at 10,000xg for 1 min. Add 1.0 mL of Standard & Sample Diluent, let it stand for 10 min and invert it gently several times. After it dissolves fully, mix it thoroughly with a pipette. This reconstitution produces a working solution of 100 ng/ml. Then make serial dilutions as needed. The recommended dilution gradient is: 100, 50, 25, 12.5, 6.25, 3.13, 1.56, and 0 ng/ml. Prepare 7 tubes, add 500 µl of Standard & Sample Diluent to each tube. Pipette 500 µl of the 100 ng/ml stock solution to the first tube and mix up to produce a 50 ng/ml working solution. Transfer 500 µl of the solution into the





other tube to form 2-fold serial dilutions of the highest standards to make the standard curve within the range of this assay.

**Sample Preparation:**

**Note:** Samples should be assayed within 7 days when stored at 4°C, otherwise aliquot and stored at -20°C (≤1 month) or -80°C (≤3 months). Avoid repeated freeze-thaw cycles

- **Serum:** Allow samples to clot for 2 hours at room temperature or overnight at 4°C before centrifugation for 20 min at 1000xg at 2–8°C. Collect the supernatant to carry out the assay. Blood collection tubes should be disposable and endotoxin free.
- **Plasma:** Collect plasma using EDTA-Na<sub>2</sub> as anticoagulant. Centrifuge samples for 15 min at 1000x g at 2–8°C within 30 min of collection. Collect the supernatant to carry out the assay. Hemolysed samples are not suitable for ELISA assay!
- **Cell lysates:** For adherent cells, gently wash the cells with moderate amount of pre-cooled PBS and dissociate the cells using trypsin. Collect the cell suspension into a centrifuge tube and centrifuge for 5 min at 1000xg. Discard the medium and wash the cells 3 times with pre-cooled PBS. For each 1x10<sup>6</sup> cells, add 150-250 µL of pre-cooled PBS to keep the cells suspended. Repeat the freeze-thaw process several times until the cells are fully lysed. Centrifuge for 10 min at 1500xg at 4°C. Remove the cell fragments, collect the supernatant for assay. Avoid repeated freeze-thaw cycles.
- **Tissue homogenates:** It is recommended to get detailed references from the literature before analyzing different tissue types. For general information, hemolysed blood may affect the results, so the tissues should be minced into small pieces and rinsed in ice-cold PBS (0.01M, pH=7.4) to remove excess blood thoroughly. Tissue pieces should be weighed and then homogenized in PBS (tissue weight (g): PBS (mL) volume=1:9) with a glass homogenizer on ice. To further break down the cells, you can sonicate the suspension with an ultrasonic cell disrupter or subject it to freeze-thaw cycles. The homogenates are then centrifuged for 5 min at 5000xg to get the supernatant.
- **Cell culture supernatant or other biological fluids:** Centrifuge samples for 20 min at 1000xg at 2- 8°C. Collect the supernatant for assay.

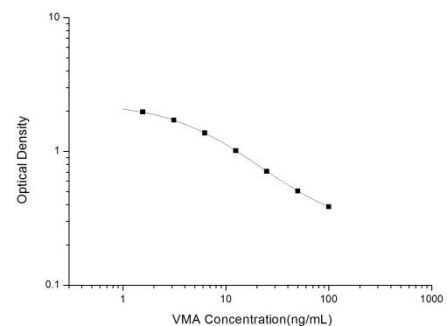
**Assay Protocol:**

**Note:** Bring all reagents and samples to room temperature 30 minutes prior to the assay. It is recommended that all standards and samples be run at least in duplicate. A standard curve must be run with each assay.

1. Add 50 µl of each **standard or samples** into appropriate wells. Immediately add 50 µL of Biotinylated Detection Ab working solution to each well. Cover the plate with the plate sealer provided in the kit and incubate for 45 min at 37°C.  
**Note:** solutions should be added to the bottom of the micro ELISA plate well, avoid touching the inside wall and bubble formation as much as possible.
2. Aspirate the solution from each well add 350 µl of **1x wash buffer** to each well. Soak for 1~2 min and aspirate or decant the solution from each well and pat it dry against clean absorbent paper. Repeat this wash step 3 times.  
**Note:** a microplate washer can be used in this step and other wash steps.
3. Add 100 µl of **HRP Conjugate working solution** to each well. Cover with the Plate sealer. Incubate for 30 min at 37°C.
- 4 Aspirate the solution from each well, repeat the wash process for five times as conducted in step 3.
- 5 Add 90 µl of **Substrate Reagent** to each well. Cover with a new plate sealer. Incubate for about 15 min at 37°C. Protect the plate from light. **Note:** the reaction time can be shortened or extended according to the actual color change, but not more than 30 min.
6. Add 50 µl of **Stop Solution** to each well. **Note:** adding the stop solution should be done in the same order as the substrate solution
7. Read the absorbance in micro plate reader set to 450 nm.

**Calculation:** Determine the average of the duplicate readings for each standard and samples then subtract the average zero standard optical density. Plot a four-parameter logistic with standard concentration on the x-axis and OD values on the y-axis. If the samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor. If the OD of the sample is under the lowest limit of the standard curve, retest the samples with appropriate dilution. The actual concentration is the concentration obtained by calculated multiplied by the dilution factor.

Typical standard curve and data is provided below for reference only. A standard curve must be run with each assay.



Concentration(ng/mL)	100	50	25	12.5	6.25	3.13	1.56	0
OD	0.386	0.506	0.711	1.012	1.373	1.714	1.971	2.318

**Precision**

Intra-assay Precision (Precision within an assay): 3 samples with low, mid-range and high level VMA were tested 20 times on one plate, respectively.

Inter-assay Precision (Precision between assays): 3 samples with low, mid-range and high level VMA were tested on 3 different plates, 20 replicates in each plate.



Sample	Intra-assay Precision			Inter-assay Precision		
	1	2	3	1	2	3
n	20	20	20	20	20	20
Mean (ng/mL)	4.60	10.90	45.90	4.80	11.80	49.40
Standard deviation	0.30	0.50	1.50	0.20	0.50	2.30
C V (%)	6.52	4.59	3.27	4.17	4.24	4.66

#### Recovery

The recovery of VMA spiked at three different levels in samples throughout the range of the assay was evaluated in various matrices.

Sample Type	Range (%)	Average Recovery (%)
Serum (n=5)	86-98	91
EDTA plasma (n=5)	93-105	99
Cell culture media (n=5)	85-99	90

#### Linearity

Samples were spiked with high concentrations of VMA and diluted with Reference Standard & Sample Diluent to produce samples with values within the range of the assay.

		Serum (n=5)	EDTA plasma(n=5)	Cell culture media(n=5)
1:2	Range (%)	94-109	97-111	97-115
	Average (%)	101	104	105
1:4	Range (%)	84-99	86-100	95-106
	Average (%)	90	92	101
1:8	Range (%)	84-96	87-103	99-114
	Average (%)	91	94	104
1:16	Range (%)	88-103	93-105	97-112
	Average (%)	95	99	103

#### VIII. Related Products:

QuickDetect™ Catecholamine (CA) (Human) ELISA Kit (E4461)  
Norepinephrine (NE) ELISA Kit (E4360)  
Dopamine (DA) ELISA Kit (K4219)  
Adrenaline/Epinephrine (EPI) ELISA Kit (E4359)

**FOR RESEARCH USE ONLY! Not to be used on humans.**