



pg/ml. Then make serial dilutions as needed. The recommended dilution gradient is as follows: 500, 250, 125, 62.5, 31.25, 15.63, 7.81, 0 pg/mL. Prepare 7 tubes; add 500 μ L of Standard and Sample Diluent to each tube. Pipette 500 μ L of the 500 pg/ml stock solution to the first tube and mix up to produce a 250 pg/ml working solution. Transfer 500 μ L of the solution into the other tube to form 2-fold serial dilutions of the highest standards to make the standard curve within the range of this assay.

VIII. Sample Preparation:

- Serum:** Allow samples to clot for 2 hours at room temperature or overnight at 2-8°C before centrifugation for 15 min at 1000xg at 2-8°C. Collect the supernatant. Blood collection tubes should be disposable and be endotoxin free.
- Plasma:** Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge samples for 15 min at 1000xg at 2-8°C within 30 min of collection. Collect the supernatant. **Hemolysed samples are not suitable for ELISA assay!**
- Cell lysates:** For adherent cells, gently wash the cells with moderate amount of pre-cooled PBS and dissociate the cells using trypsin. Collect the cell suspension into a centrifuge tube and centrifuge for 5 min at 1000xg. Discard the medium and wash the cells 3 times with pre-cooled PBS. For each 1×10^6 cells, add 150-250 μ L of pre-cooled PBS to keep the cells suspended. Repeat the freeze-thaw process several times until the cells are fully lysed. Centrifuge for 10 min at 1500xg at 2-8°C. Remove the cell fragments; collect the supernatant to carry out the assay. Avoid repeated freeze-thaw cycles.
- Tissue homogenates:** It is recommended to get detailed references from the literature before analyzing different tissue types. For general information, hemolysed blood may affect the results, so the tissues should be minced into small pieces and rinsed in ice-cold PBS (0.01M, pH=7.4) to remove excess blood thoroughly. Tissue pieces should be weighed and then homogenized in PBS (tissue weight (g): PBS (mL) volume=1:9) with a glass homogenizer on ice. To further break down the cells, sonicate the suspension with an ultrasonic cell disrupter or subject it to freeze-thaw cycles. The homogenates are then centrifuged for 5 min at 5000xg to get the supernatant.
- Cell culture supernatant or other biological fluids:** Centrifuge samples for 20 min at 1000xg at 2-8°C. Collect the supernatant to carry out the assay

IX. Assay Protocol:

Note: Bring all reagents and samples to room temperature 30 minutes prior to the assay. It is recommended that all standards and samples be run at least in duplicate. A standard curve must be run with each assay.

1. Add the **Standard working solution** to the first two columns: Each concentration of the solution is added in duplicate, to one well each, side by side (100 μ L for each well). Add the samples to the other wells (100 μ L for each well). Cover the plate with the sealer provided in the kit. Incubate for 90 min at 37°C.

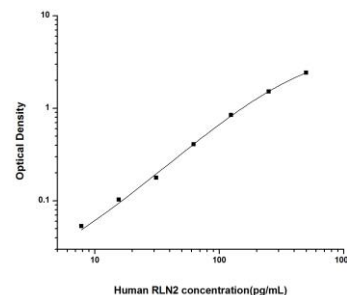
Note: solutions should be added to the bottom of the micro ELISA plate well, avoid touching the inside wall and causing foaming as much as possible

2. Remove the liquid out of each well, do not wash. Immediately add 100 μ L of **Biotinylated Detection Antibody working** solution to each well. Cover with the Plate sealer. Incubate for 1 hr at 37°C.
3. Aspirate or decant the solution from each well, add 350 μ L of **wash buffer** to each well. Soak for 1~2 min and aspirate or decant the solution from each well and pat it dry against clean absorbent paper. Repeat this wash step 3 times. Note: a microplate washer can be used in this step and other wash steps.
4. Add 100 μ L of **HRP Conjugate working solution** to each well. Cover with the Plate sealer. Incubate for 30 min at 37°C.
5. Aspirate or decant the solution from each well, repeat the wash process for five times as conducted in step 3.
6. Add 90 μ L of **Substrate Reagent** to each well. Cover with a new plate sealer. Incubate for ab out 15 min at 37°C.
 Protect the plate from light. Note: the reaction time can be shortened or extended according to the actual color change, but not more than 30 min.
7. Add 50 μ L of **Stop Solution** to each well. Note: Adding the stop solution should be done in the same order as the substrate solution.
8. Determine the optical density (OD value) of each well at once with a micro-plate reader set to 450 nm.

X. Calculation:

Average the duplicate readings for each standard and samples, then subtract the average zero standard optical density. Plot a four-parameter logistic curve on log-log graph paper, with standard concentration on the x-axis and OD values on the y-axis.

If the samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor. If the OD of the sample surpasses the upper limit of the standard curve, you should re-test it with an appropriate dilution. The actual concentration is the calculated concentration multiplied by the dilution factor.



Typical standard curve and data is provided below for reference only. A standard curve must be run with each assay.

Concentration(pg/mL)	500	250	125	62.5	31.25	15.63	7.81	0
OD	2.481	1.57	0.904	0.463	0.235	0.16	0.11	0.057
Corrected OD	2.424	1.513	0.847	0.406	0.178	0.103	0.053	-

