



# MOG (Human) ELISA Kit

(Catalog # E4963-100, 96 assays, Store at 4°C)

## I. Introduction:

Myelin oligodendrocyte glycoprotein (MOG) is a 26 kDa transmembrane glycoprotein that belongs to the immunoglobulin superfamily. MOG acts either as a cell surface receptor or a cell adhesion molecule. It is expressed exclusively by the oligodendrocytes in the central nervous system (CNS) and on the surface of myelin sheaths. It makes up about 0.05% of the myelin sheath and enables smooth electrical conduction of action potential. The extracellular IgV domain of MOG is accessible to the antibodies present in the extracellular milieu. As a result, the antibodies attack MOG and elicit a demyelinating autoantibody response. MOG antibodies are observed in disorders such as optic neuritis (ON), acute disseminated encephalomyelitis (ADEM), and transverse myelitis (TM). Hence, the detection of MOG antibodies has been implicated in the diagnosis of patients with CNS demyelination disorders. BioVision's MOG (Human) ELISA kit is based on the Sandwich principle to quantitatively measure the concentration of MOG in human serum, plasma, and other biological fluids. Test samples, Standards, and Biotinylated Detection antibody are added to the wells pre-coated with capture antibody and then washed with Wash Buffer. The HRP-Streptavidin is added and any unattached conjugates are washed off using Wash Buffer. The HRP enzymatic reaction is detected by the addition of TMB-substrate. Finally, the reaction is terminated with an acidic stop solution. The color developed is directly proportional to the concentration of MOG in the sample or standard.

## II. Features and Benefits:

- Detection range: 0.156 – 10 ng/ml
- Sensitivity: 0.094 ng/ml
- Assay Precision: Intra-Assay CV < 8% and Inter-Assay CV < 10%
- Recovery range: 85 - 105% for normal human serum and plasma samples
- This Sandwich ELISA is highly sensitive and highly specific for the detection of MOG in human samples. There is no significant cross-reactivity or interference between MOG and analogues

## III. Sample Type:

Human Serum, Plasma, Tissue lysates and other biological fluids

## IV. Kit Contents:

Components	E4963-100	Part Number	Storage Temp.
Micro ELISA plate	8 x 12 Strips	E4963-100-1	-20°C
Standard (Lyophilized) (10 ng)	2 vials	E4963-100-2	-20°C
Sample/Standard Dilution Buffer	20 ml	E4963-100-3	4°C
Biotin-labeled Antibody	120 µl	E4963-100-4	4°C (Avoid light)
Antibody Dilution Buffer	10 ml	E4963-100-5	4°C
HRP-Streptavidin Conjugate (SABC)	120 µl	E4963-100-6	4°C (Avoid light)
SABC Dilution Buffer	10 ml	E4963-100-7	4°C
TMB Substrate Solution	10 ml	E4963-100-8	4°C (Avoid light)
Stop Solution	10 ml	E4963-100-9	4°C
Wash Buffer (25X)	30 ml	E4963-100-10	4°C
Plate Sealers	5	E4963-100-11	4°C

## V. User Supplied Reagents and Equipment:

- Microplate reader capable of measuring absorbance at 450 nm
- 37°C incubator
- Precision pipettes with disposable tips
- Distilled or deionized water
- Clean eppendorf tubes for preparing standards or sample dilutions
- Absorbent paper

## VI. Storage and Handling:

The entire kit can be stored at 4°C for up to 6 months from the date of shipment.

## VII. Reagent and Sample Preparation:

**Note:** Prepare reagents within 30 minutes before the experiment

Before using the kit, spin tubes and bring down all components to the bottom of tubes

1. **Biotin-labeled Antibody working solution:** Prepare this working stock 1 hour prior to the start of the experiment. Calculate the total volume of the working solution: 0.1 ml/well x quantity of wells. Add 0.1 – 0.2 ml to the total volume. Dilute the Biotin-labeled antibody with Antibody Dilution Buffer at 1:100. Mix thoroughly.
2. **HRP-Streptavidin Conjugate (SABC):** Prepare this working stock 30 minutes prior to the start of the experiment. Calculate the total volume of the working solution: 0.1 ml/well x quantity of wells. Add 0.1 – 0.2 ml to the total volume. Dilute the SABC with SABC Dilution Buffer at 1:100. Mix thoroughly.
3. **Wash Buffer:** Dilute 25X Wash Buffer to 1X by adding 30 ml of 25X Wash Buffer and make up the volume to 750 ml with deionized/distilled water. If crystals present in the 25X Wash Buffer, warm it in water bath at 40°C. Mix it gently. The solution must be cooled to room temperature before use.
4. **Standard Preparation:**
  - Add 1 ml Sample Dilution Buffer into one Standard tube (labeled as zero tube). Keep the tube at room temperature for 10

minutes. Mix thoroughly.

- Label 7 tubes with 1/2, 1/4, 1/8, 1/16, 1/32, 1/64 and blank respectively. Add 0.3 ml of the Sample Dilution Buffer into each tube. Add 0.3 ml of the above Standard solution (from zero tube) into 1<sup>st</sup> tube and mix them thoroughly.
- Transfer 0.3 ml from 1<sup>st</sup> tube to 2<sup>nd</sup> tube and mix them thoroughly. Transfer 0.3 ml from 2<sup>nd</sup> tube to 3<sup>rd</sup> tube and mix them thoroughly, and so on. **Sample Dilution Buffer** was used for the blank control. (Note: Please use Standard Solutions within 2 hours of preparation).

## 5. Sample Preparation:

**Note:** Isolate the test samples soon after collecting, then, analyze immediately (within 2 hours). Or aliquot and store at -20°C ( $\leq$  1 month) or -80°C ( $\leq$  2 months). Avoid multiple freeze-thaw cycles. The hemolytic samples are not suitable for this assay.

- **Serum:** Place whole blood sample at room temperature for 2 hours or put it at 2 - 8°C overnight and centrifugation for 20 minutes at approximately 1000xg. Collect the supernatant and carry out the assay immediately. Blood collection tubes should be disposable, non-pyrogenic, and endotoxin free.
- **Plasma:** Collect plasma using EDTA-Na<sub>2</sub> or heparin as an anticoagulant. Centrifuge samples for 15 minutes at 1000xg at 2 - 8°C within 30 minutes of collection. Collect the supernatant and carry out the assay immediately. Avoid hemolysis, high cholesterol samples.
- **Tissue homogenates:** As hemolytic blood may affect the assay result, it is necessary to remove residual blood by washing tissue with pre-cooling PBS buffer (0.01 M, pH 7.4). Mince tissue after weighing it and homogenize it in PBS (the volume depends on the weight of the tissue. Normally, 9 ml PBS would be appropriate to 1 gram tissue pieces. Some protease inhibitors are recommended to add into the PBS) with a glass homogenizer on ice. To further break the cells, you can sonicate the suspension with an ultrasonic cell disrupter or subject it to freeze-thaw cycles. The homogenates are then centrifuged for 5 minutes at 5000xg to get the supernatant. The total protein concentration was determined by BCA kit and the total protein concentration of each pore sample should not exceed 0.3 mg.
- **Cell culture supernatant:** Centrifuge supernatant for 20 minutes at 1000xg at 2 - 8°C to remove insoluble impurity and cell debris. Collect the clear supernatant and carry out the assay immediately.
- **Cell Culture Lysate:** Commercial RIPA kits are recommended to follow the instructions provided. Generally, 0.5 ml RIPA lysis buffer would be appropriate to  $2 \times 10^6$  cells, DNA must be removed. The total protein concentration was determined by BCA kit and the total protein concentration of each pore sample should not exceed 0.3 mg.
- **Other biological fluids:** Centrifuge samples for 20 minutes at 1000xg at 4°C. Collect the supernatant and carry out the assay immediately.

**Note:** End user should estimate the concentration of the target protein in the test sample first, and select a proper dilution factor to make the diluted target protein concentration fall in the optimal detection range of the kit. **The matrix components in the sample may affect the test results. Please dilute the sample 1/2 with Sample Dilution Buffer before testing.**

## VIII. Assay Protocol:

**Note:** Bring all reagents and samples to room temperature 30 minutes prior to the assay.

It is recommended that all standards and samples be run at least in duplicate.

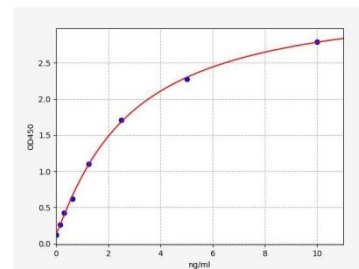
A standard curve should be run for each assay.

1. Prepare all reagents, samples (**diluted 1/2 with Sample Dilution Buffer**) and standards as instructed in section VII.
2. Wash plate 2 times with **1X Wash Buffer** before adding standard, sample (**diluted 1/2 with Sample Dilution Buffer**) and control wells.
3. Add 100  $\mu$ l of each **standards** or **samples** into appropriate wells. Cover well and incubate for 1.5 hours at 37°C.
4. Remove the cover and discard the plate content. Wash the plate 2 times with **1X Wash Buffer** without letting the wells completely dry.
5. Add 0.1 ml of **Biotin-Detection antibody** work solution into the above wells. Seal the plate and incubate at 37°C for 60 minutes.
6. Discard the solution and wash 3 times with **1X Wash Buffer**. Wash by filling each well with Wash Buffer (350  $\mu$ l) using a multi-channel pipette or autowasher. Let it soak for 1-2 minutes, and then remove all residual wash-liquid from the wells by aspiration. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Tap the plate on absorbent filter papers or other absorbent materials.
7. Add 0.1 ml of **SABC working solution** into each well, cover the plate and incubate at 37°C for 30 minutes.
8. Discard the solution and wash 5 times with **1X Wash Buffer** as step 6.
9. Add 90  $\mu$ l of **TMB substrate** into each well, cover the plate and incubate at 37 °C in dark within 10-20 minutes. (Note: The reaction time can be shortened or extended according to the actual color change, but not more than 30 minutes. The reaction can be terminated when apparent gradient appeared in standard wells).
10. Add 50  $\mu$ l of **Stop Solution** to each well. Read result at 450 nm within 20 minutes.

## IX. Calculation:

For calculation, (the relative O.D.450) = (the O.D.450 of each well) – (the O.D.450 of Zero well). The standard curve can be plotted as the relative O.D.450 of each standard solution (Y) vs. the respective concentration of the standard solution (X). The target concentration of the samples can be interpolated from the standard curve. If the samples measured were diluted, multiply the dilution factor to the concentrations from interpolation to obtain the concentration before dilution.

**Figure:** Typical Standard Curve and OD values: These standard curves are for demonstration only. A standard curve must be run with each assay.





STD. (ng/ml)	OD-1	OD-2	Average	Corrected
0	0.121	0.125	0.123	0.000
0.156	0.262	0.27	0.266	0.143
0.312	0.421	0.433	0.427	0.304
0.625	0.614	0.632	0.623	0.5
1.25	1.085	1.117	1.101	0.978
2.5	1.687	1.735	1.711	1.588
5	2.243	2.309	2.276	2.153
10	2.754	2.834	2.794	2.671

**X. Recovery:**

Matrices mentioned below were spiked with certain level of MOG and the recovery rates were calculated by comparing the measured value to the expected amount of MOG in samples

Matrix	Recovery Range (%)	Average (%)
Serum (n=5)	95-103	99
EDTA Plasma (n=5)	88-105	96
Heparin Plasma (n=5)	95-105	100

**XI. Linearity:**

Linearity of the assay kit was determined by spiking samples and their serial dilutions with appropriate concentration of MOG. The results are represented as percentage of calculated concentration to the expected value.

Sample	1:2	1:4	1:8
Serum (n=5)	86-103%	86-102%	86-100%
EDTA Plasma (n=5)	83-99%	82-100%	83-96%
Heparin Plasma (n=5)	83-99%	81-100%	86-99%

**XII. Related Products:**

- ERK2 (Human) ELISA Kit (E4315)
- Erythropoietin (Human) ELISA Kit (E4720)
- IL-17 (Human) ELISA Kit (E4628)
- AKT1/PKB (Human) ELISA Kit (K4166)
- ERK (Human) ELISA Kit (E4317)