

Caspase-8/FLICE Fluorometric Assay Kit

(Catalog #K112-25, -100, -200, -400; Store kit at -20° C)

I. Introduction:

Activation of ICE-family proteases/caspases initiates apoptosis in mammalian cells. The **FLICE/Caspase-8 Fluorometric Assay Kits** provide a simple and convenient means for assaying the activity of caspases that recognize the sequence IETD. The assay is based on detection of cleavage of substrate IETD-AFC (AFC: 7-amino-4-trifluoromethyl coumarin. Ex/Em: 400/505 nm). Upon cleavage of the substrate by FLICE or related caspases, free AFC is released, and can be quantified using a fluorometer or a fluorescence microtiter plate reader. Comparison of the fluorescence of AFC from an apoptotic sample with an uninduced control allows determination of the fold increase in FLICE activity.

II. Kit Contents:

Components	K112-25	K112-100	K112-200	K112-400	Part Number
	25 assays	100 assays	200 assays	400 assays	
Cell Lysis Buffer	25 ml	100 ml	100 ml	100 ml	K112-XX(X)-1
2X Reaction Buffer	2 ml	4 x 2 ml	16 ml	32 ml	K112-XX(X)-2
IETD-AFC (1 mM)	125 µl	0.5 ml	2 x 0.5 ml	2 x 1 ml	K112-XX(X)-3
DTT (1 M)	100 µl	0.4 ml	0.4 ml	0.4 ml	K112-XX(X)-4

III. Caspase-8 Assay Protocol:

A. General Considerations

- Aliquot enough 2X Reaction Buffer for the number of assays to be performed. Add DTT to the 2X Reaction Buffer immediately before use (10 mM final concentration: add 10 µl of 1.0 M DTT stock per 1 ml of 2X Reaction Buffer).
- After thawing, store the Cell Lysis Buffer and 2X Reaction Buffer at 4° C.
- Protect IETD-AFC from light.
- We recommend using a flat bottom, opaque, white or black 96-well plate for enhanced sensitivity.

B. Assay Procedure

1. Induce apoptosis in cells by desired method. Concurrently incubate a control culture *without* induction.
2. Count cells and pellet 1-5 x 10⁶ cells or use 50-200 µg cell lysates if protein concentration has been measured.
3. Resuspend cells in 50 µl of chilled Cell Lysis Buffer. Incubate cells on ice for 10 minutes.
4. Add 50 µl of 2X Reaction Buffer (containing 10 mM DTT) to each sample. Add 5 µl of the 1 mM IETD-AFC substrate (50 µM final concentration). Incubate at 37° C for 1-2 hours.
5. Read samples in a fluorometer equipped with a 400-nm excitation filter and 505-nm emission filter. For a plate-reading set-up, transfer the samples to a 96-well plate. You may also perform the entire assay directly in a 96-well plate.
Fold-increase in FLICE activity can be determined by comparing these results with the level of the uninduced control.

IV. Storage and Stability:

Store kit at -20° C (Store Cell Lysis Buffer and 2X Reaction Buffer at 4° C after opening). All reagents are stable for 6 months under proper storage conditions.

V. Related Products:

Apoptosis Detection Kits & Reagents

- Annexin V Kits & Bulk Reagents
- Caspase Assay Kits & Reagents
- Mitochondrial Apoptosis Kits & Reagents
- Nuclear Apoptosis Kits & Reagents
- Apoptosis Inducers and Apoptosis siRNA Vectors

Cell Fractionation System

- Mitochondria/Cytosol Fractionation Kit
- Nuclear/Cytosol Fractionation Kit
- Membrane Protein Extraction Kit
- Mammalian Cell Extraction Kit
- FractionPREP Fractionation System

Cell Proliferation & Senescence

- Quick Cell Proliferation Assay Kit
- Senescence Detection Kit
- High Throughput Apoptosis/Cell Viability Assay Kits
- LDH-Cytotoxicity Assay Kit
- Bioluminescence Cytotoxicity Assay Kit
- Live/Dead Cell Staining Kit

Cell Damage & Repair

- HDAC & HAT Fluorometric & Colorimetric Assays & Drug Discovery Kits
- DNA Damage Quantification Kit
- Glutathione & Nitric Oxide Fluorometric & Colorimetric Assay Kits

Signal Transduction

- cAMP & cGMP Assay Kits
- Akt & JNK Activity Assay Kits

Adipocyte & Lipid Transfer

- Recombinant Adiponectin, Survivin, & Leptin
- CETP & PLTP Activity Assay & Drug Discovery Kits
- Cholesterol Quantification Kit

Molecular Biology & Reporter Assays

- siRNA Vectors
- Cloning Insert Quick Screening Kit
- Mitochondrial & Genomic DNA Isolation Kits
- 5 Minutes DNA Ligation Kit
- 20 Minutes Gel Staining/Destaining Kit

Growth Factors and Cytokines

- Adiponectin/Resistin/Leptin and their Antibodies
- Recombinant Protein A and Protein G
- Recombinant Cytokines and Growth Factors
- Recombinant Complement C5a

Problems	Cause	Solution
GENERAL TROUBLESHOOTING GUIDE FOR CASPASE COLORIMETRIC AND FLUOROMETRIC KITS:		
		; buffer and incubate as described in the datasheet
	<ul style="list-style-type: none"> Experiment was not performed at optimal time after apoptosis induction Plate read at incorrect wavelength Old DTT used 	<ul style="list-style-type: none"> Perform a time-course induction experiment for apoptosis Check the wavelength listed in the datasheet and the filter settings of the instrument Always use freshly thawed DTT in the cell lysis buffer
High Background	<ul style="list-style-type: none"> Increased amount of cell lysate used Increased amounts of components added due to incorrect pipetting Incubation of cell samples for extended periods Use of expired kit or improperly stored reagents Contaminated cells 	<ul style="list-style-type: none"> Refer to datasheet and use the suggested cell number to prepare lysates Use calibrated pipettes Refer to datasheet and incubate for exact times Always check the expiry date and store the individual components appropriately Check for bacterial/ yeast/ mycoplasma contamination
Lower signal levels	<ul style="list-style-type: none"> Cells did not initiate apoptosis Very few cells used for analysis Use of samples stored for a long time Incorrect setting of the equipment used to read samples Allowing the reagents to sit for extended times on ice 	<ul style="list-style-type: none"> Determine the time-point for initiation of apoptosis after induction (time-course experiment) Refer to datasheet for appropriate cell number Use fresh samples or aliquot and store and use within one month for the assay Refer to datasheet and use the recommended filter setting Always thaw and prepare fresh reaction mix before use
Samples with erratic readings	<ul style="list-style-type: none"> Uneven number of cells seeded in the wells Samples prepared in a different buffer Adherent cells dislodged and lost at the time of experiment Cell/ tissue samples were not completely homogenized Samples used after multiple freeze-thaw cycles Presence of interfering substance in the sample Use of old or inappropriately stored samples 	<ul style="list-style-type: none"> Seed only equal number of healthy cells (correct passage number) Use the cell lysis buffer provided in the kit Perform experiment gently and in duplicates/triplicates; apoptotic cells may become floaters Use Dounce homogenizer (increase the number of strokes); observe efficiency of lysis under microscope Aliquot and freeze samples, if needed to use multiple times Troubleshoot as needed Use fresh samples or store at correct temperatures until use
Unanticipated results	<ul style="list-style-type: none"> Measured at incorrect wavelength Cell samples contain interfering substances 	<ul style="list-style-type: none"> Check the equipment and the filter setting Troubleshoot if it interferes with the kit (run proper controls)
General issues	<ul style="list-style-type: none"> Improperly thawed components Incorrect incubation times or temperatures Incorrect volumes used Air bubbles formed in the well/tube Substituting reagents from older kits/ lots Use of a different 96-well plate 	<ul style="list-style-type: none"> Thaw all components completely and mix gently before use Refer to datasheet & verify the correct incubation times and temperatures Use calibrated pipettes and aliquot correctly Pipette gently against the wall of the well/tubes Use fresh components from the same kit Fluorescence: Black plates; Absorbance: Clear plates
Note: The most probable cause is listed under each section. Causes may overlap with other sections.		