



- 1 vial (100 µg) of exosome standards (lyophilized), from human urine (number of particles/ml 1×10^{10}).
- Exosome standards: The remaining reconstituted standard stock solution should be aliquoted into polypropylene vials (preferably low binding) and stored at -20°C for up to one month or at -80°C for up to six months. Strictly avoid repeated freeze-and-thaw cycles.
- Store opened and diluted reagents at 4°C up to 12 months if unopened.

VIII. ExoQuant™ Assay Protocol:

- 1. Human Urine sample preparation:** Centrifuge at 350g for 10 min at room temperature and save the supernatant.
- 2. Exosome isolation from Human Urine:**

| Fluid | Minimum volume required | Volume Suggested |
|-------|-------------------------|------------------|
| Urine | 5 ml | 8 ml – 20 ml |

- Add ExoPure™ Reagent to your sample in ratio 1/4 (i.e. 5 ml of urine + 1.2 ml of ExoPure™).
 - Mix well by pipetting and inverting tube.
 - Incubate on ice for 1 hr.
 - Centrifuge 20 min at 10,000g (centrifuge can be performed at 4°C or at RT).
 - Discard the supernatant.
 - Centrifuge for 2 min at 1500g to eliminate entirely the supernatant.
 - Resuspend the pellet in 100 µl* of 1X PBS. * Volume of resuspension can be defined by the user.
 - Resuspended exosomes can be used for analysis or stored at -20°C
- 3. Lyophilized Exosome Standard reconstitution:**
- Reconstitute lyophilized exosome standard by adding 100 µl of deionized water to get a final concentration of 1 µg/µl.
 - Resuspend exosomes pipetting the solution up and down 10-15 times, avoiding bubbles.
 - Vortex the reconstituted standard for 60 secs. Briefly centrifuge the tubes containing the standard to ensure that the solution is collected at the bottom of the tube. Pipette the solution up and down 10 times, avoiding the introduction of bubbles. After this step, the standard is ready to use.
 - Use 5 µl of reconstituted Exosome Standard for each reaction.
- 4. Exosome binding onto latex FACS-Beads:**
- Dilute 5 times Sample buffer 5x with PBS 1x (Sample buffer 1x).
 - It is recommended to prepare the complex Exosome-Beads (Exo-Beads) in one single tube, then to divide in single reactions before the antibody incubation.
 - Latex FACS-Beads are ready to use. Resuspend well FACS-Beads prior to use by vortexing or pipetting several times.
 - For each reaction mix together 5 µl of Exosome Standards and 5 µl of FACS-beads in an eppendorf tube (preferably low binding). Mix well by pipetting 5-6 times. **Example: if you want to run 10 reactions, mix into the same eppendorf low binding tube 50 µl of Exosome Standards and 50 µl of FACS-Beads.**
 - For exosome isolated by EXO-Prep, mix 5 µl of FACS-Beads with the volume of resuspended exosomes suggested (volumes are indicative only; the user should define the appropriate volumes on the base of exosome yield).
 - **Urine: 50 µl - 100 µl /reaction**
 - Incubate for 15 min at room temperature (RT).
 - Add 0.7 ml of PBS 1x and incubate in rotator or shaker for 2 hrs at RT or overnight (ON) at 4°C.
 - Centrifuge the complex Exosomes-Beads (Exo-Beads) for 5 min at 4500 g at 4°C and discard the supernatant.
- 5. Antibody Incubation:**
- Prepare the Washing buffer (not provided in the kit) diluting 2% of FBS (or FCS) in PBS 1x (consider that you need 8 ml of washing buffer for each reaction). Alternatively, if you don't have FBS or FCS, prepare the Washing buffer diluting 0.5% of BSA in PBS 1X. Keep on ice.
 - Resuspend the Exo-Beads in 100 µl of Sample buffer 1x for each reaction. Example: if you are running 10 reactions resuspend Exo-Beads in 1 ml of Sample buffer.
 - Prepare the FACS tubes (not provided in the kit), one tube for each reaction.
 - Divide the Exo-Beads resuspended in sample buffer in each FACS tube, pipetting 100 µl of suspension in each tube.
 - Add primary antibody in ratio 1:200 (0.5 µl per each FACS tube)*.
 - *If other primary antibodies are used the correct dilution must be defined by the user.**
 - Incubate 2 hours at 4°C in the dark. For negative control can be used PE or FITC-anti Mouse IgG1 isotype or FITC or PE anti-Rabbit IgG1. **(isotype controls not provided in the kit)**. Otherwise incubate control samples with secondary Abs only (either leave in ice or directly add sec Abs).
 - Add 4 ml of prepared Washing buffer to each FACS tube.
 - Centrifuge 5 min at 4000 g and discard the supernatant.
 - Resuspend beads pellet into the FACS tubes in 100 µl of Sample buffer 1x.
 - Add secondary antibody in ratio 1:1000. (mix 38 µl of Sample buffer 1x with 2 µl of secondary antibody; add 2 µl of the received solution per each FACS tube to obtain the right dilution of antibody).
 - Incubate for 1 hr in the dark at 4°C.
 - Add 4 ml of Washing buffer in each FACS tube, centrifuge for 5 min at 4000g 4°C. Discard supernatant by pouring it out. Vortex what has remained inside the FACS tubes. If not use immediately, put in the dark at 4°C.
 - Add 500 µl of Washing buffer per FACS tube.
 - Analyze the samples.
 - Read 10,000 events from the gated first population.
 - Alexa 488 is read in FL1 channel (green)

