

Caspase-4 Colorimetric Assay Kit

(Catalog #K127-25, -100, -200, -400; Store kit at -20°C)

I. Introduction:

Activation of ICE-family proteases/caspases initiates apoptosis in mammalian cells. The **Caspase-4 Colorimetric Assay Kit** provides a simple and convenient means for assaying the activity of caspases that recognize the sequence LEVD. The assay is based on spectrophotometric detection of the chromophore *p*-nitroanilide (*p*NA) after cleavage from the labeled substrate LEVD-*p*NA. The *p*NA light emission can be quantified using a spectrophotometer or a microtiter plate reader at 400- or 405-nm. Comparison of the absorbance of *p*NA from an apoptotic sample with an uninduced control allows determination of the fold increase in LEVD-dependent Caspase activity.

II. Kit Contents:

Components	K127-25	K127-100	K127-200	K127-400	Part Number
	25 assays	100 assays	200 assays	400 assays	
Cell Lysis Buffer	25 ml	100 ml	100 ml	100 ml	K127-XX(X)-1
2X Reaction Buffer	2 ml	4 x 2 ml	16 ml	32 ml	K127-XX(X)-2
LEVD- <i>p</i> NA (4 mM)	125 µl	0.5 ml	2 x 0.5 ml	2 x 1 ml	K127-XX(X)-3
DTT (1 M)	100 µl	0.4 ml	0.4 ml	0.4 ml	K127-XX(X)-4
Dilution Buffer	25 ml	100 ml	200 ml	400 ml	K127-XX(X)-5

III. Caspase-4 Assay Protocol:

A. Reagent Preparations

Aliquot enough 2X Reaction Buffer for the number of assays to be performed. Add DTT to the 2X Reaction Buffer immediately before use (10 mM final concentration: Add 10 µl of the 1.0 M DTT per 1 ml of 2X Reaction Buffer).

B. Assay Procedure

1. Induce apoptosis in cells by desired method. Concurrently incubate a control culture *without* induction.
2. Count cells and pellet 5 x 10⁶ cells.
3. Resuspend cells in 50 µl of chilled Cell Lysis Buffer and incubate on ice for 10 minutes.
4. Centrifuge for 1 min in a microcentrifuge (10,000 x g).
5. Transfer supernatant (cytosolic extract) to a fresh tube and put on ice.
6. Assay protein concentration.
7. Dilute 200-300 µg protein to 50 µl Cell Lysis Buffer for each assay.
8. Add 50 µl of 2X Reaction Buffer (containing 10 mM DTT) to each sample. Add 5 µl of the 4 mM LEVD-*p*NA substrate (200 µM final conc.). Incubate at 37°C for 1-2 hour.
9. Read samples at 400- or 405-nm in a microtiter plate reader, or spectrophotometer using a 100-µl micro quartz cuvette (Sigma), or dilute sample to 1 ml with Dilution Buffer and using regular cuvette (note: Dilution of the samples proportionally decreases the reading). You may also perform the entire assay in a 96-well plate.

Fold-increase in LEVD-dependent caspase activity can be determined by comparing readings of induced samples with the level of uninduced control.

Note: Background reading from cell lysates and buffers should be subtracted from the readings of both induced and the uninduced samples before calculating fold increase in caspase activity.

IV. Storage and Stability:

- Store kit at -20°C (Store Cell Lysis Buffer, 2X Reaction Buffer, and Dilution Buffer at 4°C after opening). Protect LEVD-*p*NA from light.
- All reagents are stable for 1 year under proper storage conditions.

V. Related Products:

Apoptosis Detection Kits & Reagents

- Annexin V Kits & Bulk Reagents
- Caspase Assay Kits & Reagents
- Mitochondrial Apoptosis Kits & Reagents
- Nuclear Apoptosis Kits & Reagents
- Apoptosis Inducers and Apoptosis siRNA Vectors

Cell Fractionation System

- Mitochondria/Cytosol Fractionation Kit
- Nuclear/Cytosol Fractionation Kit
- Membrane Protein Extraction Kit
- Cytosol/Particulate Rapid Separation Kit
- Mammalian Cell Extraction Kit
- FractionPREP Fractionation System

Cell Proliferation & Senescence

- Quick Cell Proliferation Assay Kit
- Senescence Detection Kit
- High Throughput Apoptosis/Cell Viability Assay Kits
- LDH-Cytotoxicity Assay Kit
- Bioluminescence Cytotoxicity Assay Kit
- Live/Dead Cell Staining Kit

Cell Damage & Repair

- HDAC & HAT Fluorometric & Colorimetric Assays & Drug Discovery Kits
- DNA Damage Quantification Kit
- Glutathione & Nitric Oxide Fluorometric & Colorimetric Assay Kits

Signal Transduction

- cAMP & cGMP Assay Kits
- Akt & JNK Activity Assay Kits
- Beta-Secretase Activity Assay Kit

Adipocyte & Lipid Transfer

- Recombinant Adiponectin, Survivin, & Leptin
- CETP & PLTP Activity Assay & Drug Discovery Kits
- Cholesterol Quantification Kit

Molecular Biology & Reporter Assays

- siRNA Expression Vectors
- Cloning Insert Quick Screening Kit
- Mitochondrial & Genomic DNA Isolation Kits
- 5 minutes DNA Ligation Kit

Growth Factors and Cytokines

GENERAL TROUBLESHOOTING GUIDE FOR CASPASE COLORIMETRIC AND FLUOROMETRIC KITS

Problems	Cause	Solution
Assay not working	<ul style="list-style-type: none"> Cells did not lyse completely Experiment was not performed at optimal time after apoptosis induction Plate read at incorrect wavelength Old DTT used 	<ul style="list-style-type: none"> Resuspend the cell pellet in the lysis buffer and incubate as described in the datasheet Perform a time-course induction experiment for apoptosis Check the wavelength listed in the datasheet and the filter settings of the instrument Always use freshly thawed DTT in the cell lysis buffer
High Background	<ul style="list-style-type: none"> Increased amount of cell lysate used Increased amounts of components added due to incorrect pipetting Incubation of cell samples for extended periods Use of expired kit or improperly stored reagents Contaminated cells 	<ul style="list-style-type: none"> Refer to datasheet and use the suggested cell number to prepare lysates Use calibrated pipettes Refer to datasheet and incubate for exact times Always check the expiry date and store the individual components appropriately Check for bacteria/ yeast/ mycoplasma contamination
Lower signal levels	<ul style="list-style-type: none"> Cells did not initiate apoptosis Very few cells used for analysis Use of samples stored for a long time Incorrect setting of the equipment used to read samples Allowing the reagents to sit for extended times on ice 	<ul style="list-style-type: none"> Determine the time-point for initiation of apoptosis after induction (time-course experiment) Refer to datasheet for appropriate cell number Use fresh samples or aliquot and store and use within one month for the assay Refer to datasheet and use the recommended filter setting Always thaw and prepare fresh reaction mix before use
Samples with erratic readings	<ul style="list-style-type: none"> Uneven number of cells seeded in the wells Samples prepared in a different buffer Adherent cells dislodged and lost at the time of experiment Cell/ tissue samples were not completely homogenized Samples used after multiple freeze-thaw cycles Presence of interfering substance in the sample Use of old or inappropriately stored samples 	<ul style="list-style-type: none"> Seed only equal number of healthy cells (correct passage number) Use the cell lysis buffer provided in the kit Perform experiment gently and in duplicates/triplicates; apoptotic cells may become floaters Use Dounce homogenizer (increase the number of strokes); observe efficiency of lysis under microscope Aliquot and freeze samples, if needed to use multiple times Troubleshoot as needed Use fresh samples or store at correct temperatures until use
Unanticipated results	<ul style="list-style-type: none"> Measured at incorrect wavelength Cell samples contain interfering substances 	<ul style="list-style-type: none"> Check the equipment and the filter setting Troubleshoot if it interferes with the kit (run proper controls)
General issues	<ul style="list-style-type: none"> Improperly thawed components Incorrect incubation times or temperatures Incorrect volumes used Air bubbles formed in the well/tube Substituting reagents from older kits/ lots Use of a different 96-well plate 	<ul style="list-style-type: none"> Thaw all components completely and mix gently before use Refer to datasheet & verify the correct incubation times and temperatures Use calibrated pipettes and aliquot correctly Pipette gently against the wall of the well/tubes Use fresh components from the same kit Fluorescence: Black plates; Absorbance: Clear plates
Note# The most probable cause is listed under each section. Causes may overlap with other sections.		