



01/19

Periodic Acid Schiff Diastase Staining Kit

(Cat# K1433-30, -250; PAS; Store at Multiple Temperatures)

I. Introduction:

The Periodic Acid Schiff (PAS) Diastase Staining Kit is intended for use in histological demonstration of lymphocytes and mucopolysaccharides. The a-Amylase digestion step acts on glycogen to break it into smaller sugars that are then washed off the tissue section allowing visual comparison of digested and undigested slides. The PAS reaction in tissue sections is useful for the demonstration of mucopolysaccharides.

PAS Positive Material: Magenta; Nuclei: Blue

- II. Application:
 - Histological applications, IHC-P
 - For in vitro diagnostic use

III. Sample Type:

- Any well fixed paraffin embedded tissue sections (3-5 microns)
- Control Tissue: Liver
- IV. Kit Contents:

Components	K1433-30	K1433-250	Part Number	Storage Temperature
Alpha-Amylase Solution (1%)	30 ml	250 ml	K1433-XX-1	2-8°C
Periodic Acid Solution	30 ml	250 ml	K1433-XX-2	2-8°C
Schiff's Solution	30 ml	250 ml	K1433-XX-3	2-8°C
Hematoxylin, Mayer's	30 ml	2 x 125 ml	K1433-XX-4	RT
Bluing Reagent	30 ml	2 x 125 ml	K1433-XX-5	RT

V. User Supplied Reagents and Equipment:

- Distilled water
- Coplin jars
- Forceps
- · Absolute alcohol
- Synthetic resin

VI. Shipment and Storage:

All the reagents are shipped at room temperature and stored at multiple temperatures.

VII. Reagent Preparation:

- Do not use if reagents become cloudy.
- Do not use past expiration date.
- Use caution when handling reagents.
- Non-Sterile

VIII. Procedure (Standard):

Deparaffinize two identical sections if necessary and hydrate to distilled water.

- If sections are Zenker-fixed, remove mercuric chloride crystals using iodine and clear with sodium thiosulfate. Rinse in running tap water.
 Apply Alpha-Amylase Solution (1%) to one slide and incubate for 10-30 minutes at room temperature.
- 4. Rinse in 2 changes of distilled water. Note: The remainder of this procedure is performed on both the "digested" and "undigested" slides.
- 5. Apply Periodic Acid Solution (1%) to tissue section and incubate for 5 minutes.
- 6. Rinse slide in 4 changes of distilled water.
- 7. Apply Schiff's Solution to tissue section and incubate for 10-20 minutes.
- 8. Rinse slide in warm running tap water for 2 minutes.
- 9. Rinse slide in distilled water.
- 10. Apply Hematoxylin, Mayer's (Lillie's Modification) to tissue section and incubate for 1 minute.
- 11. Rinse in running tap water for 1 minute followed by 2 changes of distilled water.
- 12. Apply Bluing Reagent for 5 seconds and rinse in distilled water.
- 13. Dehydrate through graded alcohols.
- 14. Clear, and mount in synthetic resin.

FOR RESEARCH USE ONLY! Not to be used on humans.

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