



# **Plasmid DNA Extraction Kit**

(Catalog# K1445-100; 100 tests; Storage at Multiple Temperatures)

#### I. Introduction:

Plasmid DNA extraction and purification kit with proprietary separation and buffer systems, allows extracting and purifying plasmid DNA from 2.0~5.0 mL of bacteria culture. The typical yield is 10~20 µg of high quality plasmid DNA and the typical OD260/280 between 1.75 to 1.85. Purified plasmid DNA can be directly used for a variety of molecular biology applications such as enzymatic digestion, sequencing and transformation.

The kit will work with a 48 well round bottom plates if a special magnetic frame is used. The kit can also be used with a variety of automatic nucleic acid extraction instruments or workstations.

#### II. Applications:

• Extraction and purification of plasmid DNA from bacteria culture. Purified DNA can be used for enzymatic digestion, sequencing, transformation etc.

#### III. Sample Type:

Bacterial cultures

# IV. Kit Contents:

Component	K1445-100	Part Number
Magnetic Beads	5 mL	K1445-100-1
RNase A Solution	55 µl	K1445-100-2
Suspension Solution	22 ml	K1445-100-3
Lysis Solution	22 ml	K1445-100-4
Neutralization Solution	40 ml	K1445-100-5
Wash Solution*	38 ml	K1445-100-6
Elution Buffer	10 ml	K1445-100-7

\* Add 25 ml of Isopropanol to Wash Solution\* (K1445-100-6) before use.

#### V. User Supplied Reagents and Equipment:

- 80% Ethanol in water.
- · Magnetic racks compatible with vials.
- Isopropanol (ACS grade).

#### VI. Storage Conditions and Reagent Preparation:

Magnetic beads should be stored at 2-8°C but other kit reagents need to be stored at room temperature. Lysis solution may turn cloudy if stored in the cold room. To clear it up place the bottle in a water bath at 37°C.

# Reagent Preparation:

Wash solution\* (K1445-100-6): Dilute the solution by adding 25 ml of Isopropanol before using.

#### VII. Assay Protocol:

- 1. Preparation. Before the first use, add all the RNase A solution into the Suspension Solution.
- 2. Harvest cells. Spin 1.5-5 mL of overnight grown bacterial cells at 12,000 rpm for 2 min.
- 3. Re-suspend the cells. Remove culture media completely. Add 200 µl of suspension solution (with RNase A), then vortex to ensure complete suspension of cell. Transfer cells to a clean Eppendorf tube.
- 4. Lyse the cells. Add 200 µl of Lysis solution and mix by inverting the tube for 6-8 times. Incubate for 2 min. Do NOT vortex.
- 5. Neutralize the solution. Add 350 µl of Neutralization solution and mix by inverting the tube for 8-10 times. Do NOT vortex.
- 6. Spin the tube at 12,000 rpm for 15 min at 4°C.

7. Transfer the solution to a clean Eppendorf tube, then add 50 µl of magnetic beads, mix well and incubate 3-5 min at RT. Put Eppendorf tube onto the magnet rack for 20 seconds.

- 8. Remove solution by holding the magnet rack upside down or by pipetting.
- 9. Wash the beads with 500 µl of Wash Solution and then repeat Step 8.
- 10. Wash the beads with 500  $\mu l$  of 80% ethanol twice repeating Step 8.
- 11. Dry the beads at 55°C for 8 min leaving the tube open. Do not over-dry the beads.
- 12. Elute the DNA from beads with 50-100 µl of elution buffer, incubate for at least 2 min and then vortex at full speed for 1

min. alternatively, incubation at 60°C for 2 min may improve the recovery for DNA larger than 10 kb.

- 13. Remove beads by using magnet rack, pipette DNA out and transfer to a clean tube.
- 14. Store purified DNA at -20°C for long-term storage.

### VIII. Related Products:

- Plasmid Miniprep Kit (# K529)
- Plasmid Miniprep Kit I (# K1312)
- Plasmid Midi Kit II (# K1315)
- Plasmid Midi Kit I (# K1314)
- Plasmid Miniprep Kit II (# K1313)

## FOR RESEARCH USE ONLY! Not to be used on humans.