

2. Enzyme Solution Preparation: Mix enough reagents for the number of assays to be performed. Prepare a 100-fold dilution of the Human ODC1 (e.g. Mix 2 μl of Human ODC1 with 198 μl ODC1 Assay Buffer. 50% glycerol solution is viscous. Handle human ODC1 solution carefully. For each well, prepare 40 μl ODC1 Enzyme Solution:

ODC1 Assay Buffer	20 μl
Diluted Human ODC1	20 μl

Mix and add 40 μl of the ODC1 enzyme solution to Sample, Inhibitor Control, Enzyme Control and Solvent Control wells ([S], [IC], [EC] and [SolC]). Add 40 μl of Assay Buffer into Background Control and Sample control ([BC] and [SC]) well(s). Mix well, and incubate the plate for 5 min at 37 $^{\circ}\text{C}$.

Note: Do not store unused diluted DFMO solutions.

3. Reaction Mix: Mix enough reagents for the number of assays to be performed. For each well, prepare 50 μl Mix containing:

Reaction Mix

ODC1 Assay Buffer	34 μl
ODC1 Substrate	2 μl
ODC1 Converter	2 μl
ODC1 Cofactor	10 μl
ODC1 Enzyme Mix	2 μl

Mix and add 50 μl of the Reaction Mix to all wells ([S], [EC], [IC], [SC], [BC], [SolC]). Mix well.

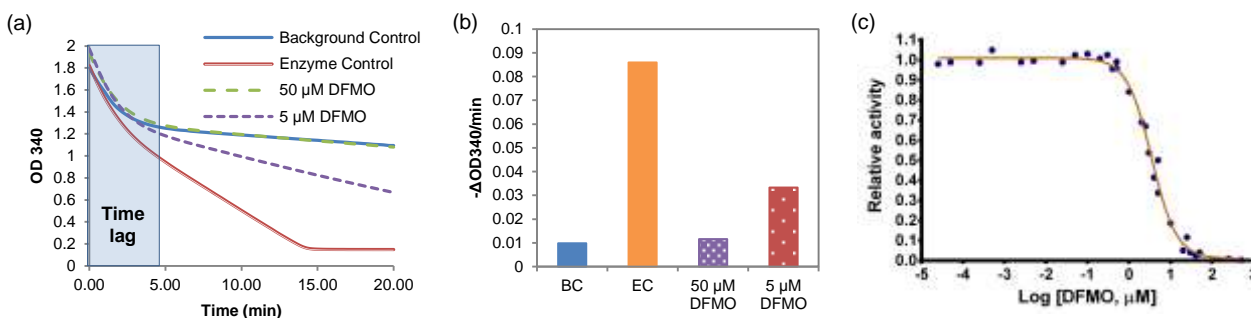
4. Measurement: Immediately, measure absorbance (OD: 340 nm) in a microplate reader in kinetic mode at 37 $^{\circ}\text{C}$ for 20-30 min, taking measurement readings every 20 seconds.

5. Calculation: Take the linear portion of the kinetic curve $\Delta\text{OD} = \text{OD}_{\text{end}} - \text{OD}_{\text{initial}}$ and divide by reaction time ($\Delta t = t_{\text{end}} - t_{\text{initial}}$) to get the rate of individual well (note ΔOD rates would be negative). Subtract the Background Control [BC] rate from all readings to obtain activity for each reading (if Sample Control [SC] is higher than Background Control [BC] for a given test compound, subtract its rate from the signal of that particular sample only). Set the activity of Enzyme Control [EC] as 100% (in case Solvent Control is significantly different from EC, replace with [SolC] values in the formulas below). Calculate % Inhibition or % Relative Activity of the test inhibitors as follows:

$$\% \text{ Inhibition} = \frac{\text{Activity of EC} - \text{Activity of S}}{\text{Activity of EC}} \times 100$$

$$\% \text{ Relative Activity} = \frac{\text{Activity of S}}{\text{Activity of EC}} \times 100$$

Note: A slow kinetic response in the reactions (Lag phase) may be observed. **Do not use the first 3-5 minutes of the reaction curves for the estimation of ODC activity.**



Figures: (a) ODC1 was incubated with different amounts of DFMO for 5 minutes at 37 $^{\circ}\text{C}$. Kinetic curves were taken for 20 minutes at 37 $^{\circ}\text{C}$. (b) Decrease Rates ($\text{OD}_{340}/\text{min}$) of ODC1 incubated with different amounts of DFMO. (c) Inhibition of ODC1 enzyme activity by DL-2-difluoromethylornithine (DFMO). IC_{50} of DL-DFMO: $3.52 \pm 0.19 \mu\text{M}$. Assay was performed following the kit protocol.

VII. RELATED PRODUCTS:

Ornithine Decarboxylase, Human Recombinant (P1342)
Ornithine Assay Kit (Kxxx)
Arginase I Inhibitor Screening Assay Kit (K567)
ARG1, Human Recombinant (P1032)
Methionine Assay Kit (Fluorometric) (K442)
Urea Colorimetric Assay Kit (K375)

Total Polyamine Assay Kit (K475)
L-Arginine (Colorimetric) Assay Kit (K749)
Diamine Oxidase Activity Assay Kit (K496)
Arginase 2, Human Recombinant (P1308)
Arginase activity Colorimetric Assay Kit (K755)
Urea Colorimetric Assay Kit II (K376)

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