

Note:

For Unknown Samples, we suggest testing several doses of your Sample to make sure the readings are within the Standard Curve range.

2. Standard Curve Preparation: Add 0, 4, 8, 12, 16 & 20 μl of the 100 mM NADH Oxidase Standard into a series of wells in 96-well clear plate to generate 0, 8, 16, 24, 32, and 40 nmol/well of NADH Oxidase Standard. Adjust the volume of all Standard wells to 50 μl with NADH Oxidase Assay Buffer.

3. NADH Oxidase Positive Control: Add 10-15 μl of the reconstituted NADH Oxidase Positive Control into the desired well(s). Adjust the volume to 50 μl /well with NADH Oxidase Assay Buffer.

4. Reaction Mix Preparation: Mix enough reagents for the number of assays to be performed. For each well, prepare 50 μl Reaction Mix containing:

	<u>Reaction Mix</u>	<u>Standard Mix</u>
NADH Oxidase Assay Buffer	32 μl	34 μl
NADH Oxidase Enzyme Mix	2 μl	2 μl
NADH Oxidase Substrate I	4 μl	4 μl
NADH Oxidase Substrate II	10 μl	10 μl
NADH Oxidase Probe	2 μl	---

Mix well. Add 50 μl of Reaction Mix into Positive Control and Sample wells and 50 μl of Standard Mix into Standard wells respectively.

5. Measurement: Measure the absorbance (OD_{600 nm}) in kinetic mode for 30 min at 25 °C. **Note:** Incubation time depends on the NADH Oxidase activity in the Samples. We recommend measuring the absorbance in kinetic mode and choosing any two time points (T₁ & T₂) in the linear range to calculate the NADH Activity. The NADH Oxidase Standard Curve can be read in Endpoint mode (at the end of 30 min incubation).

6. Calculation: Subtract 0 Standard reading from all Standard readings and plot the NADH Oxidase Standard Curve. Subtract the Sample readings from the Blank readings to get the corrected Sample reading. Apply the corrected Sample reading to the NADH Oxidase Standard Curve to get 'B' nmol of product generated during the reaction time ($\Delta T = T_2 - T_1$).

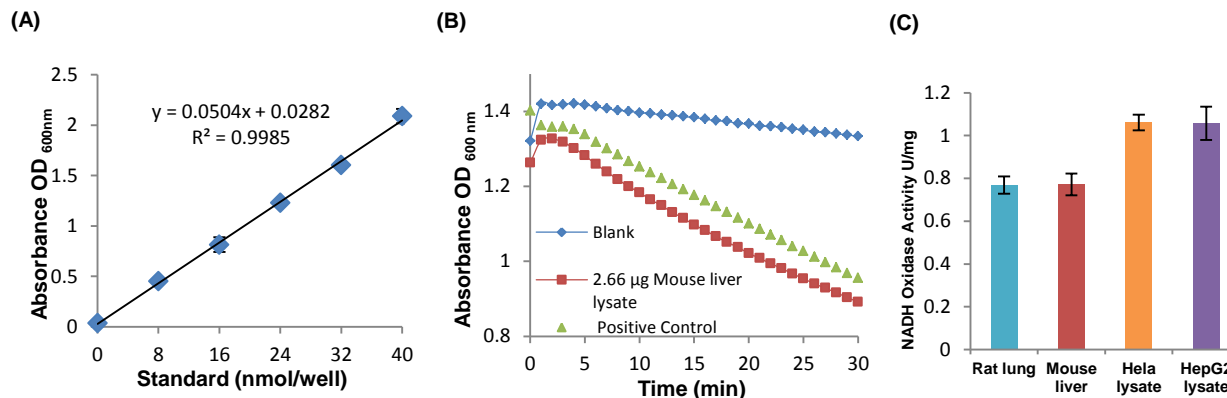
To determine the activity of NADH Oxidase in Sample(s), use the following equation:

$$\text{Sample NADH Oxidase Activity} = \frac{B}{\Delta T \times P} = \text{nmol/min/mg} = \text{mU/mg}$$

Where: **B** = Product amount from the NADH Oxidase Standard Curve (nmol)

ΔT = Difference between T₂ and T₁ (min)

P = Amount of protein in the Sample (mg)



Figures: (A) NADH Oxidase Standard Curve. (B) Reaction kinetics of recombinant NADH Oxidase (Positive Control) and Mouse liver lysate. (C) NADH Oxidase activity detected in Rat lung lysate (2.8 μg), Mouse liver lysate (2.66 μg), HeLa cell lysate (6 μg) and HepG2 cell lysate (3 μg). Assays were performed according to the kit protocol.

VIII. Related Products:

- PicoProbe™ LDH-Cytotoxicity Fluorometric Assay Kit (# K314)
- PicoProbe™ NADH Fluorometric Assay Kit (# K338)
- Lactate Dehydrogenase A Inhibitor Screening Kit (# K492)
- Glucose Dehydrogenase Activity Colorimetric Assay Kit (# K786)

- NAD/NADH Quantitation Colorimetric Kit (# K337)
- PicoProbe™ NADPH Quantitation Fluorometric Assay Kit (# K349)
- Lactate Dehydrogenase Colorimetric Assay Kit (# K726)
- EZScreen™ NAD+/NADH Colorimetric Assay Kit (384-well) (# K958)

FOR RESEARCH USE ONLY! Not to be used on humans.