



2. Ammonium Chloride Standard Curve Preparation: Dilute the 100 mM Ammonium Chloride Standard to 1 mM Ammonium Chloride Standard by adding 10 µl of 100 mM Ammonium Chloride Standard to 990 µl of PAL Assay Buffer. Further, dilute the 1 mM Ammonium Chloride Standard to 200 µM Ammonium Chloride Standard working solution by adding 200 µl of 1 mM Ammonium Chloride Standard to 800 µl of PAL Assay Buffer. Add 0, 2, 4, 6, 8 and 10 µl of 200 µM Ammonium Chloride Standard working solution to each wells to generate 0, 0.4, 0.8, 1.2, 1.6 and 2.0 nmoles of Ammonium Chloride Standard/well, respectively. Adjust the volume of each well to 100 µl with PAL Assay Buffer.

Note: Ammonia present in the air can result in a high background.

3. Reaction Mix Preparation: Mix enough reagents for the number of assays to be performed. For each well, depending on the contents, either prepare a total of 50 µl Reaction Mix or 50 µl Sample Background Mix containing:

	<u>Reaction Mix</u>	<u>Sample Background Mix</u>
PAL Assay Buffer	48 µl	50 µl
PAL Substrate	2 µl	-

Add 50 µl Reaction Mix to each well containing Sample, Substrate Background Control, and Positive Control and mix well. Add 50 µl Sample Background Mix to Sample Background Control well(s) and mix well. Cover the plate with a Microplate Sealing Film and incubate at 37°C for 30 min.

4. Developer B Preparation: While the plate is being incubated, prepare Developer B by adding 11 µl of 2-Mercaptoethanol to 1989 µl of 200-proof Ethanol. Mix well and keep on ice while in use.

Note: Always prepare Developer B fresh before each experiment and keep on ice, when in use.

5. Developer Mix Preparation: Prepare enough Developer Mix for the number of assays to be performed.

	<u>Developer Mix</u>
PAL Assay Buffer	86 µl
Developer A	7 µl
Developer B	7 µl

Following the 30 min incubation of the plate from step 3, unseal the plate and add 100 µl of Developer Mix to all wells including Sample, Sample Background Control, Substrate Background Control, Positive Control and Ammonium Chloride Standards. Mix well and re-seal the plate.

6. Measurement: Incubate the plate at 37°C for another 30 min. After the 30 min incubation, remove the Microplate Sealing Film and record the fluorescence at Ex/Em = 410/470 nm in end point mode.

7. Calculation: Subtract the 0 Standard reading from all Standard readings and Sample Background Control reading from Sample readings, respectively. Plot the Ammonium Chloride Standard Curve. If the Substrate Background Control is higher than the Sample Background Control, subtract the Substrate Background Control from the Sample readings instead. Apply the corrected Sample readings to the Ammonium Chloride Standard Curve to get the value of B nmol of Ammonium ions in the sample.

$$\text{Sample Phenylalanine Ammonia-Lyase Activity} = (B / \Delta t) \times D \text{ (nmol/min)} = mU$$

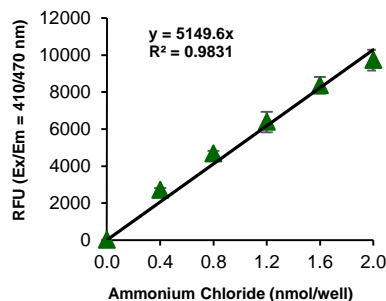
Where, **B** = Amount of Ammonium ions in the sample (nmol)

Δt = Reaction time (i.e. 30 min)

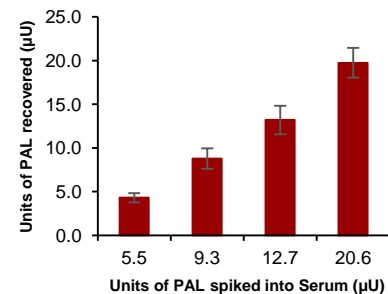
D = Dilution factor (D= 1, for undiluted samples)

Unit Definition: One unit of Phenylalanine Ammonia-Lyase is the amount of enzyme that produces 1.0 µmol of Ammonium ions per minute at pH 8.0 at 37°C.

a)



b)



Figures. (a). Ammonium Chloride Standard Curve. **(b).** Pooled normal human male AB serum was spiked with 5.5, 9.3, 12.7 and 20.6 µU of PAL respectively. The corresponding units of PAL recovered were 4.3, 8.8, 13.2 and 19.7 µU (recovery rates ranging from 78.4-103.7%). The data shown is the average of three replicates where experiments were performed according to the kit protocol.

VIII. Related Products:

Phenylalanine Assay Kit (Colorimetric) (K481)

Phenylalanine Fluorometric Assay Kit (K572)

Potassium (Serum) Detection Assay Kit (Fluorometric) (K940)

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