



b. Organic solvents used to prepare the test inhibitor stock solutions may impact BChE activity. In order to determine the effect of solvent on the BChE activity, we recommend preparing a parallel Solvent Control [SC] well with the same final concentration of the solvent used to solubilize the test inhibitor(s). If the signal obtained in the Solvent Control [SC] well is significantly different from the [EC] well, use the signal for the SC well instead of the EC well.

2. BChE Enzyme Solution Preparation: Prepare a 10-fold dilution of the reconstituted BChE Enzyme by adding 5 µl of BChE Enzyme to 45 µl of BChE Assay Buffer and mix well. Add 5 µl of diluted BChE Enzyme to each well containing Sample [S], Inhibitor Control [IC], Solvent Control [SC] and Enzyme Control [EC]. Add 5 µl of BChE Assay Buffer to the Background Control [BC] well. Adjust the volume of all wells [S, IC, SC, EC and BC] to **80 µl/well** using BChE Assay Buffer. **Mix well and incubate at RT for 30 min, protected from light.**
Note: Discard any unused, diluted BChE Enzyme after use.

3. Reaction Mix Preparation: Prepare a 12-fold dilution of the BChE Substrate (*i.e.* mix 5 µl of BChE Substrate with 55 µl of BChE Assay Buffer). Further dilute the diluted BChE Substrate at a 1:10 dilution. Mix enough reagents for the number of assays to be performed. For each well, prepare 20 µl Reaction Mix containing:

	<u>Reaction Mix</u>
Diluted BChE Substrate	10 µl
Probe Mix	5 µl
BChE Assay Buffer	5 µl

Mix well and add 20 µl Reaction Mix to Sample [S], Inhibitor Control [IC], Enzyme Control [EC], Solvent Control [SC], and Background Control [BC] wells and mix well. The total reaction volume in each well will be 100 µl.

4. Measurement: Measure the absorbance immediately in kinetic mode for 30-60 min at 412 nm at RT. Choose any two time points (t_1 and t_2) in the linear range of the plot and obtain the corresponding OD values for Sample (OD_1 and OD_2).

5. Calculation: Calculate the slope of Sample [S], Enzyme Control [EC], Solvent Control [SC] and Background Control [BC] by dividing the $\Delta OD_{412\text{nm}}$ ($OD_2 - OD_1$) values over reaction time Δt ($t_2 - t_1$). Subtract the slope of the Background Control [BC] values from the [S], [EC] and [SC] values. If the [SC] slope is significantly different from the [EC], use the [SC] value instead to calculate the effect of the test inhibitor(s).

$$\% \text{ Relative Inhibition} = \frac{\text{Slope of [EC]} - \text{Slope of [S]}}{\text{Slope of [EC]}} \times 100$$

$$\% \text{ Relative Activity} = \frac{\text{Slope of [S]}}{\text{Slope of [EC]}} \times 100$$

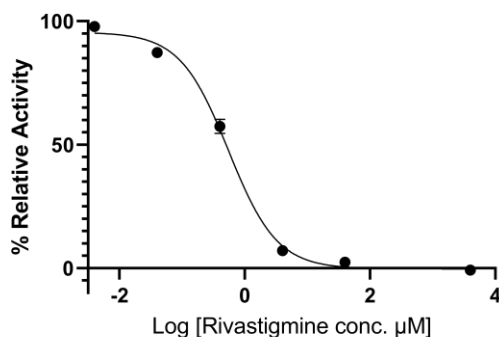


Figure: Inhibition of BChE activity by Rivastigmine. IC_{50} of Rivastigmine was calculated to be 550.9 ± 120.2 nM. Assay was performed following the kit protocol.

VII. Related Products:

- Rivastigmine tartrate (B3093)
- Acetylcholinesterase Inhibitor Screening Kit (Colorimetric) (K197)
- Acetylcholinesterase Activity Colorimetric Assay Kit (K764)
- Butyrylcholinesterase Activity Kit (K516)
- Cholinesterase Activity Assay Kit (K975)
- BChE Antibody (Center) (6724)
- Phosphate Buffered Saline (PBS) (2113)

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