

# HDAC8 Activity Fluorometric Assay Kit

(Catalog #K348-100; 100 assays; Store kit at -80°C)

## I. Introduction:

Histone deacetylases (HDACs) represent a large family of enzymes identified as key regulators of nucleosomal histone acetylation, a major event that controls eukaryotic gene transcription and are classified into three groups. HDACs are transcriptional repressors with crucial roles in mammalian development. They are believed to be involved in important biological activities, such as cell differentiation, proliferation, apoptosis, and senescence. In BioVision's HDAC8 activity Kit, HDAC8 deacetylates the substrate (R-H-K(Ac)-K(Ac)-AFC) followed by an enzymatic cleavage to release the AFC molecule, which can be detected fluorometrically (Ex/Em = 380/500 nm). The kit provides a rapid, simple, sensitive, and reliable test, which is also suitable for high throughput measurement of HDAC8 activity in purified, immunoprecipitated and recombinant or genetically modified HDAC-8 samples.

## II. Kit Contents:

Components	100 assays	Cap Color	Part Number
HDAC8 Assay Buffer	25 ml	WM	K348-100-1
HDAC8 Substrate	200 µl	Red	K348-100-2
HDAC8 Positive Control	20 µl	Green	K348-100-3
AFC Standard (1 mM)	100 µl	Yellow	K348-100-4
Developer	1 ml	Orange	K348-100-5
Trichostatin A (100 µM)	200 µl	Blue	K348-100-6

## III. Storage and Handling:

Remove the vial of HDAC8 and store the kit at -20°C, protect from light. Allow Assay Buffer to warm to room temperature before use. Briefly centrifuge vials before opening. Read the entire protocol before performing the assay.

**HDAC8 Positive Control:** Aliquot and store at -80°C. Use within 2 months.

**HDAC8 Substrate:** Upon thawing, aliquot and store at -20°C, use within 2 months.

**Developer and Trichostatin A:** Store at -20°C; use within 2 months.

**HDAC8 Assay Buffer:** Store at 4°C

## IV. HDAC-8 Activity Assay Protocol:

- Standard Curve Preparation:** Prepare a 10 µM AFC (7-amino-4-trifluoromethyl coumarin) Standard by mixing 10 µl of the 1 mM AFC Standard with 990 µl of dH<sub>2</sub>O. Add 0, 2, 4, 6, 8, 10 µl 10 µM AFC into individual wells in a 96-well plate and adjust the volume to 100 µl/well with HDAC8 Assay Buffer to generate 0, 20, 40, 60, 80, 100 pmol/well of AFC Standard. Read fluorometrically at Em/Ex = 380/500 nm after mixing.
- Sample Preparations:** Prepare test samples of up to 50 µl/well with HDAC8 Assay Buffer in a 96-well plate in replicates. Add 2 µl HDAC-8 Assay Buffer to one sample replicate and 2 µl HDAC8 Inhibitor (Trichostatin A) to another sample replicate as the sample background control. Mix well and incubate for 10 min at 37 °C. We suggest testing several doses of your sample to ensure the readings are within the standard curve linear range. Use 1 - 2 µl HDAC8 as a positive control and adjust volume to 50 µl with HDAC8 Assay Buffer.

## 3. Substrate Preparation:

- 36 µl HDAC8 Assay Buffer
- 2 µl HDAC8 Substrate

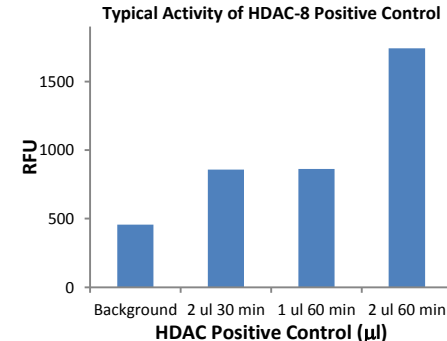
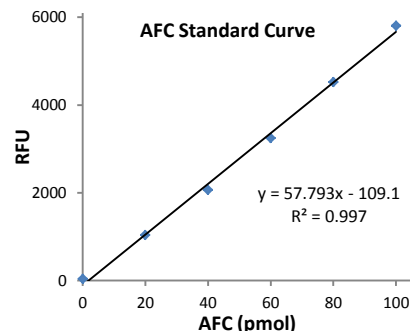
Add 38 µl substrate solution into each well (**Do Not add to Standard Curve wells**). Mix well. Incubate at 37°C for 60 min.

- Developer:** Add 10 µl into each well (**Do Not add to Standard Curve wells**). Mix well, incubate another 5 min at 37°C.
- Measurement:** Read Ex/Em = 380/500 nm R<sub>B</sub> and R<sub>S</sub> for each background well and sample well, respectively.
- Calculation:** Plot the AFC Standard Curve. The RFU of fluorescence generated is Δ RFU = R<sub>S</sub> - R<sub>B</sub>. Apply the Δ RFU to the standard curve to get B pmol of AFC:

$$\text{Activity} = \frac{B}{(60) \times V} \times \text{Sample Dilution Factor} = \text{pmol/min/ml} = \mu\text{M/ml}$$

Where: B is the AFC amount from the Standard Curve (in pmol).  
60 is the sample/substrate incubation time (in min).  
V is the sample volume added into the reaction well (in ml).

**Unit Definition:** One unit is defined as the amount of HDAC8 able to generate 1.0 µmol of AFC per minute at 37°C when incubated with the HDAC8 Substrate (R-H-K(Ac)-K(Ac)-AFC).



## RELATED PRODUCTS:

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| <ul style="list-style-type: none"> <li>HDAC-1 thru HDAC11 Antibodies</li> <li>DiscoveryPak™ HDAC Inhibitor Set</li> <li>HDAC Activity Assay Fluorometric Kit</li> <li>HAT (P/CAF) Active Human Recombinant</li> <li>Trichostatin A</li> <li>HDAC8, human recombinant</li> <li>HDAC3 Inhibitor Screening Assay Kit</li> </ul> | <ul style="list-style-type: none"> <li>HDAC Family Antibody Set</li> <li>HDAC Activity Assay Colorimetric Kit</li> <li>HAT Activity Assay Colorimetric Kit</li> <li>HAT-1 thru HAT-3 Antibodies</li> <li>HDAC8 Inhibitor Screening Kit</li> <li>HDAC3 Activity Assay Kit</li> <li>HDAC3, human recombinant</li> </ul> |
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**FOR RESEARCH USE ONLY! Not to be used on humans.**

**GENERAL TROUBLESHOOTING GUIDE:**

Problems	Cause	Solution
Assay not working	<ul style="list-style-type: none"> <li>• Use of ice-cold assay buffer</li> <li>• Omission of a step in the protocol</li> <li>• Plate read at incorrect wavelength</li> <li>• Use of a different 96-well plate</li> </ul>	<ul style="list-style-type: none"> <li>• Assay buffer must be at room temperature</li> <li>• Refer and follow the data sheet precisely</li> <li>• Check the wavelength in the data sheet and the filter settings of the instrument</li> <li>• Fluorescence: Black plates (clear bottoms) ; Luminescence: White plates ; Colorimeters: Clear plates</li> </ul>
Samples with erratic readings	<ul style="list-style-type: none"> <li>• Use of an incompatible sample type</li> <li>• Samples prepared in a different buffer</li> <li>• Cell/ tissue samples were not completely homogenized</li> <li>• Samples used after multiple free-thaw cycles</li> <li>• Presence of interfering substance in the sample</li> <li>• Use of old or inappropriately stored samples</li> </ul>	<ul style="list-style-type: none"> <li>• Refer data sheet for details about incompatible samples</li> <li>• Use the assay buffer provided in the kit or refer data sheet for instructions</li> <li>• Use Dounce homogenizer (increase the number of strokes); observe for lysis under microscope</li> <li>• Aliquot and freeze samples if needed to use multiple times</li> <li>• Troubleshoot if needed</li> <li>• Use fresh samples or store at correct temperatures until use</li> </ul>
Lower/ Higher readings in Samples and Standards	<ul style="list-style-type: none"> <li>• Improperly thawed components</li> <li>• Use of expired kit or improperly stored reagents</li> <li>• Allowing the reagents to sit for extended times on ice</li> <li>• Incorrect incubation times or temperatures</li> <li>• Incorrect volumes used</li> </ul>	<ul style="list-style-type: none"> <li>• Thaw all components completely and mix gently before use</li> <li>• Always check the expiry date and store the components appropriately</li> <li>• Always thaw and prepare fresh reaction mix before use</li> <li>• Refer datasheet &amp; verify correct incubation times and temperatures</li> <li>• Use calibrated pipettes and aliquot correctly</li> </ul>
Readings do not follow a linear pattern for Standard curve	<ul style="list-style-type: none"> <li>• Use of partially thawed components</li> <li>• Pipetting errors in the standard</li> <li>• Pipetting errors in the reaction mix</li> <li>• Air bubbles formed in well</li> <li>• Standard stock is at an incorrect concentration</li> <li>• Calculation errors</li> <li>• Substituting reagents from older kits/ lots</li> </ul>	<ul style="list-style-type: none"> <li>• Thaw and resuspend all components before preparing the reaction mix</li> <li>• Avoid pipetting small volumes</li> <li>• Prepare a master reaction mix whenever possible</li> <li>• Pipette gently against the wall of the tubes</li> <li>• Always refer the dilutions in the data sheet</li> <li>• Recheck calculations after referring the data sheet</li> <li>• Use fresh components from the same kit</li> </ul>
Unanticipated results	<ul style="list-style-type: none"> <li>• Measured at incorrect wavelength</li> <li>• Samples contain interfering substances</li> <li>• Use of incompatible sample type</li> <li>• Sample readings above/below the linear range</li> </ul>	<ul style="list-style-type: none"> <li>• Check the equipment and the filter setting</li> <li>• Troubleshoot if it interferes with the kit</li> <li>• Refer data sheet to check if sample is compatible with the kit or optimization is needed</li> <li>• Concentrate/ Dilute sample so as to be in the linear range</li> </ul>

**Note:** The most probable list of causes is under each problem section. Causes/ Solutions may overlap with other problems.