



	Reaction Mix	Background Control Mix
ADP Assay Buffer	44 μ l	46 μ l
ADP Enzyme Mix	2 μ l	---
Developer	2 μ l	2 μ l
Probe	2 μ l	2 μ l

Add 50 μ l of the Reaction Mix to each well containing the Standard and test samples. Mix well.

Note: For samples having high NADH, add 50 μ l of Background Control Mix to sample background control well(s). Mix well.

4. Measurement: Incubate for 20 min at 37°C and measure OD_{450nm}.

5. Calculation: Subtract 0 Standard reading from all readings. Plot the ADP Standard Curve. **Note:** For samples having high NADH, correct sample background by subtracting the value derived from the background control from all sample readings. Apply the corrected sample reading to the Standard Curve to get B nmol of ADP amount in the sample.

$$\text{Sample ADP concentration} = B/V \times \text{Dilution Factor} = \text{nmol/ml} = \mu\text{M}$$

Where: **B** is the ADP amount from the Standard Curve (nmol).

V is the sample volume added into the reaction well (ml).

ADP molecular weight: 501.32 g/mole.

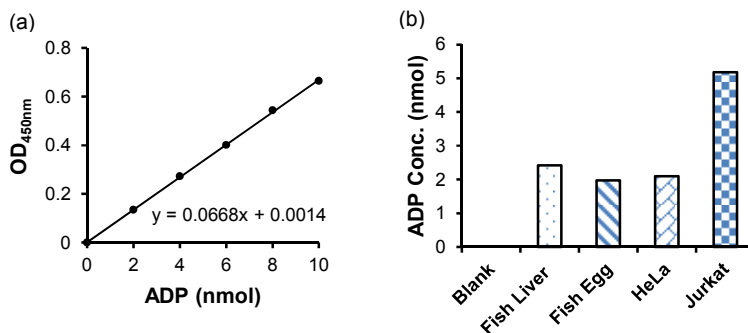


Figure 1. ADP Standard Curve [a]. Measurement of ADP in fish liver (100 μ g), fish egg (200 μ g) & HeLa & Jurkat cell lysate (100 μ g) [b]. Assays were performed following kit protocol.

IX. RELATED PRODUCTS:

ADP Colorimetric/Fluorometric Assay Kit
ApoSENSOR™ ADP/ATP Ratio Assay Kit
ATP Colorimetric/Fluorometric Assay Kit
StayBrite™ Highly Stable ATP Assay Kit
ApoSENSOR™ Cell Viability Assay Kit

Live/Dead Cell Staining Kit
StayBrite™ Highly Stable Luciferase/Luciferin Reagent
NAD/NADH Quantification Kit
NADP/NADPH Quantification Kit
Cell Proliferation Assay Kits

FOR RESEARCH USE ONLY! Not to be used on humans.