

TACE Inhibitor Screening Assay Kit (Fluorometric)

(Catalog #K366-100; 100 assays; Store kit at -20°C)

I. Introduction: The TACE (tumor necrosis factor- α -converting enzyme), also called ADAM metallopeptidase domain 17 (ADAM17), is a 70-kDa enzyme that belongs to the ADAM protein family of disintegrins and metalloproteases. TACE is believed to be involved in the processing of tumor necrosis factor alpha (TNF- α) at the surface of the cell, and from within the intracellular membranes of the trans-Golgi network. This process, which is also known as 'shedding', involves the cleavage and release of a soluble ectodomain from membrane-bound pro-proteins (such as pro-TNF- α), and is of known physiological importance. In BioVision's TACE inhibitor screening Kit, TACE hydrolyzes the specific FRET substrate to release the quenched fluorescent group, which can be detected fluorometrically at Ex/Em = 318/449 nm. In the presence of the potent TACE inhibitor, the hydrolyzation of substrate will be impeded. The kit provides a rapid, simple, sensitive and reliable test suitable for high-throughput screening of TACE inhibitors and can be modified to check the relative TACE activity. Inhibitor Control GM6001 is included to compare the efficacy of test inhibitors.

II. Kit Contents:

Components	K366-100	Cap Code	Part Number
TACE Assay Buffer	25 ml	WM	K366-100-1
TACE Substrate	0.2 ml	Red	K366-100-2
TACE Enzyme (20 μ g)	1 vial	Green	K366-100-3
Inhibitor Control (0.1 mM GM6001)	20 μ l	Purple	K366-100-4

III. Storage and Handling:

Store kit at -20°C, protected from light. Warm Assay Buffer to room temperature before use. Briefly centrifuge all small vials before opening. Read the entire protocol before performing the assay.

IV. Reagent Preparation:

TACE Enzyme: Dissolve the TACE enzyme with 220 μ l Assay Buffer. Aliquot and store the stock solution at -80°C (preferable) or -20°C. Avoid repeated freeze/thaw cycles. Use within one week.

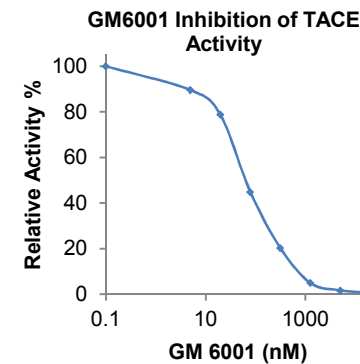
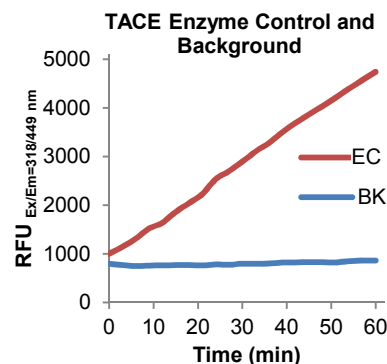
V. TACE Inhibitor Screening Assay Protocol:

- Enzyme preparation:** For each well, prepare 50 μ l TACE enzyme solution.
 - 48 μ l Assay Buffer
 - 2 μ l TACE enzyme
- Screen compounds, inhibitor control and blank control preparations:** Dissolve candidate inhibitors into proper solvent. Dilute to 4X the desired test concentration with Assay Buffer. For GM6001 Inhibitor Control, dilute GM6001 1:24 with Assay buffer. Add 25 μ l diluted test inhibitors, Inhibitor Control or Assay Buffer into TACE enzyme wells as sample screen [S], Inhibitor Control (GM6001), or Enzyme Control [EC] (no inhibitor). Mix well, and incubate for 5 minutes at 37°C.
- Substrate preparation:** For each well, prepare a total 25 μ l substrate solution
 - 23 μ l Assay Buffer
 - 2 μ l Substrate
 Mix, add 25 μ l substrate solution into each well. Mix well.

- Measurement:** Read fluorescence (R₁) at Ex/Em = 318/449 nm. Incubate the reaction at 37°C for 30 min, protected from light and measure again fluorescence (R₂) at Ex/Em = 318/449 nm.
- Calculation:** The RFU of fluorescence generated by hydrolyzation of substrate is Δ RFU = R₂ - R₁. Set the Δ RFU of Enzyme Control [EC] as 100%, and calculate the relative % inhibition of the test inhibitors as:

$$\% \text{ Inhibition} = \frac{\Delta\text{RFU of EC} - \Delta\text{RFU of S}}{\Delta\text{RFU of EC}} \times 100\%$$

Note: It is recommended to read kinetically to choose the R₁ and R₂ at linear range.



RELATED PRODUCTS:

- TACE, human recombinant
- MMP-1, human recombinant
- MMP-2, human recombinant
- MMP-3, human recombinant
- MMP-8, human recombinant
- MMP-9, human recombinant
- Pro-MMP-13, human recombinant
- MMP-3 inhibitor screen kit
- MMP-3 assay kit
- MMP-17 Antibody
- GM6001

- TACE Antibody
- MMP-1 Antibody
- MMP-2 Antibody
- MMP-3 Antibody
- MMP-8 Antibody
- MMP-9 Antibody
- MMP-13 Antibody
- MMP-11 Antibody
- MMP-12 Antibody
- MMP19 Antibody

FOR RESEARCH USE ONLY! Not to be used on humans.

GENERAL TROUBLESHOOTING GUIDE:

Problems	Cause	Solution
Assay not working	<ul style="list-style-type: none"> • Use of ice-cold assay buffer • Omission of a step in the protocol • Plate read at incorrect wavelength • Use of a different 96-well plate 	<ul style="list-style-type: none"> • Assay buffer must be at room temperature • Refer and follow the data sheet precisely • Check the wavelength in the data sheet and the filter settings of the instrument • Fluorescence: Black plates (clear bottoms) ; Luminescence: White plates ; Colorimeters: Clear plates
Lower/ Higher readings in Samples and Standards	<ul style="list-style-type: none"> • Improperly thawed components • Use of expired kit or improperly stored reagents • Allowing the reagents to sit for extended times on ice • Incorrect incubation times or temperatures • Incorrect volumes used 	<ul style="list-style-type: none"> • Thaw all components completely and mix gently before use • Always check the expiry date and store the components appropriately • Always thaw and prepare fresh reaction mix before use • Refer datasheet & verify correct incubation times and temperatures • Use calibrated pipettes and aliquot correctly
Readings do not follow a linear pattern for Standard curve	<ul style="list-style-type: none"> • Use of partially thawed components • Pipetting errors in the standard • Pipetting errors in the reaction mix • Air bubbles formed in well • Standard stock is at an incorrect concentration • Calculation errors • Substituting reagents from older kits/ lots 	<ul style="list-style-type: none"> • Thaw and resuspend all components before preparing the reaction mix • Avoid pipetting small volumes • Prepare a master reaction mix whenever possible • Pipette gently against the wall of the tubes • Always refer the dilutions in the data sheet • Recheck calculations after referring the data sheet • Use fresh components from the same kit
Unanticipated results	<ul style="list-style-type: none"> • Measured at incorrect wavelength • Samples contain interfering substances • Use of incompatible sample type • Sample readings above/below the linear range 	<ul style="list-style-type: none"> • Check the equipment and the filter setting • Troubleshoot if it interferes with the kit • Refer data sheet to check if sample is compatible with the kit or optimization is needed • Concentrate/ Dilute sample so as to be in the linear range
<p>Note: The most probable list of causes is under each problem section. Causes/ Solutions may overlap with other problems.</p>		