



4. Use 50 µl per well for the assay.

Note: Dilution factor: 5

• **Urine**

1. Centrifuge 0.5 ml of urine at 10,000 x g for 5 min and recover the supernatant.
2. Dilute the supernatant 5 fold with Sample Diluent. For example, mix 100 µl of urine with 400 µl of Sample Diluent.
3. Use 50 µl per well for the assay.

Note: Dilution factor: 5

• **Milk**

1. Add 20 µl of Extraction Solution to 1 ml of milk and vortex well.
2. Centrifuge the sample at 10,000 x g for 20 min at 4°C and recover the clear supernatant.
3. Dilute the supernatant 5 fold with Sample Diluent. For example, mix 100 µl of the supernatant with 400 µl of Sample Diluent.
4. Use 50 µl per well for the assay.

Note: Dilution factor: 5

• **Tissue (pork, liver, chicken, fish and shrimp)**

1. Weigh 1 g of the tissue sample. Mix the tissue with 1 ml of water and 20 µl of Extraction Solution. Homogenize and vortex for 5 min.
2. Centrifuge the sample at 10,000 x g for 20 min at 4°C and recover the supernatant.
3. Dilute the supernatant 5 fold with Sample Diluent. For example, mix 100 µl of the supernatant with 400 µl of Sample Diluent
4. Use 50 µl per well for the assay.

Note: Dilution factor: 5

IX. Gentamicin ELISA Assay Protocol:

Notes: It is recommended that all standards and samples should be run at least in duplicate.

Standard curves must be run each time an assay is performed.

1. Prepare all reagents, standards and samples as indicated in sections VII and VIII respectively.
2. Add 50 µl of Standards or Samples per well. Then add 50 µl of conjugate working solution and 50 µl of Antibody to the above wells.
3. Cover the microtiter plate with the plate sealer and mix well. Incubate the plate at 25°C for 30 min.
4. After incubation time, aspirate all reagents and wash each well 4 times: add 250 µl of 1X Wash Buffer and incubate for 30 seconds. Repeat this step three more times. *Complete removal of 1X Wash buffer is essential for accurate results.*
5. Add 100 µl of TMB Substrate to each well. Tap or shake the plate to ensure complete mixing.
6. Check the OD at 650 nm for the well containing no gentamicin (S0). When it reaches 0.8 – 1.0, add 50 µl of Stop Solution to each well and gently tap the plate to ensure thorough mixing.
7. Measure the OD at 450 nm for the standards and samples immediately.

X. Calculation:

The mean values of relative absorbance for the standards or samples are divided by the absorbance value of the zero-standard (S0) and multiplied by 100%. The zero-standard is set to 100% and the relative absorbances of the standards and samples (A) are expressed as percentages.

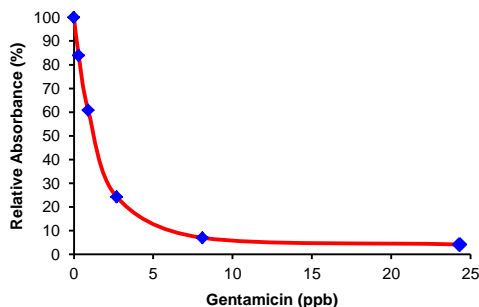
$$\text{Relative Absorbance (\%)} = A/A_0 \times 100\%$$

A: The average absorbance of the standard or sample

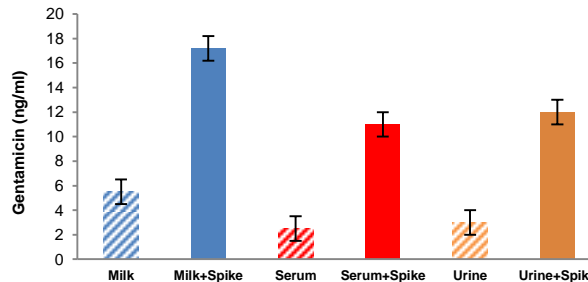
A₀: The average absorbance of the zero standards

The Gentamicin Standard Curve is done by plotting the relative absorbance of the standards vs. gentamicin concentrations (ppb). The concentration of Gentamicin of each sample (ppb), which can be read from the calibration curve, is multiplied by the corresponding dilution factor.

A.



B.



Figures. A. Gentamicin standard curve (*This standard curve is for demonstration only. A standard curve must be run with each assay*). B. Spike recovery experiment: Milk, human serum and urine samples were assayed with and without spike (10 ng/ml). Experiments showed 80-100% recovery.

XI. RELATED PRODUCTS:

Ampicillin sodium (Cat. No. 2484-5G, 25G, 100G)
Colistin Sulfate (Cat. No. 9696-1G, 5G)

Carbencillin disodium (Cat. No. 2485-1G, 5G, 10G)
Penicillin G sodium (Cat. No. 2503-100, 500)

FOR RESEARCH USE ONLY! Not to be used on humans.