



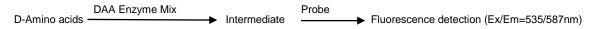
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# **Total D-Amino Acid Assay Kit (Fluorometric)**

(Catalog #K445-100; 100 assays; Store at -20°C)

## I. Introduction:

D-Amino acids are present in both eukaryotes and prokaryotes. D-Amino acids act as a component of the cell wall in prokaryotes and a neurological regulator in eukaryotes. In mammals, D-Amino acids play an important role in physiological functions and their levels are regulated by D-Amino acid oxidase in the central nervous systems. Some important D-Amino acids include D-serine, D-aspartate and D-alanine. D-serine acts as a modulator of *N*-Methyl-D-aspartate receptor (NMDA receptor, NMDAR), while D-aspartate and D-alanine have been found to be elevated in the white and the gray matter respectively in patients' brains suffering Alzheimer's disease. The abnormal levels of D-Amino acids could be used as a marker of neurological diseases: total D-Amino acid concentration in cerebrospinal fluid (CSF) has been shown to be higher in patients with Alzheimer's disease when compared to samples from healthy donors. BioVision's Total D-Amino Acid Assay Kit (Fluorometric) provides a quick, specific and easy method for measuring total D-Amino acid concentrations in a wide variety of samples. In this assay, D-Amino acids are converted into an intermediate by the DAA enzyme mix that will further react with a probe to produce a strong fluorescence signal (Ex/Em= 535/587nm). The kit is simple to use, sensitive and high-throughput adaptable and can detect as low as 1.9 µM of D-Amino acids in biological samples.



## II. Applications:

- · Measurement of total D-Amino acids concentrations in biological samples
- Analysis of relationship between D-Amino acids in physiological conditions including schizophrenia, Alzheimer disease, etc.

# III. Sample Type:

- Biological fluids: cerebrospinal fluid, etc.
- · Animal tissues: brain, etc.

#### IV. Kit Contents:

Components	K445-100	Cap Code	Part Number
DAA Assay Buffer	25 ml	WM	K445-100-1
DAA Cofactor	1 vial	Brown	K445-100-2
DAA Enzyme Mix	1 vial	Blue	K445-100-3
DAA Developer Mix	1 vial	Green	K445-100-4
DAA Probe	200 µl	Red	K445-100-5
DAA Standard	1 vial	Yellow	K445-100-6

## V. User Supplied Reagents and Equipment:

- 96-well clear plate with flat bottom
- Multi-well spectrophotometer (ELISA reader)
- Dounce Tissue Homogenizer (Cat. #1998)

## VI. Storage Conditions and Reagent Preparation:

Store kit at -20°C, protected from light. Briefly centrifuge small vials prior to opening. Read entire protocol before performing the assay.

- DAA Assay Buffer: Warm to room temperature before use. Store at 4°C or -20°C.
- DAA Cofactor: Reconstitute with 220 μl dH<sub>2</sub>O. Keep on ice while in use. Store at -20°C. Avoid light. Use within two months.
- DAA Enzyme Mix, DAA Developer Mix: Reconstitute each vial with 220 µl DAA Assay buffer. Aliquot and store at -20°C. Keep on ice while in use. Avoid freeze and thaw. Use within two months.
- DAA Probe (in DMSO): Ready to use as supplied. Warm to room temperature before use. Store at -20°C. Avoid excessive exposure to
  light.
- D-Amino acid Standard: Reconstitute with 110 μl of dH<sub>2</sub>O to make a 10 mM stock solution. Store at -20°C, stable for 5 freeze/thaw cycles.

## VII. D-Amino Acid Oxidase Assay Protocol:

1. Sample Preparation: For tissue samples: Rapidly homogenize tissue (~10 mg) in 100 μl ice cold DAAO Assay Buffer with Dounce Tissue Homogenizer (Cat. #1998), and keep on ice for 10 min. Centrifuge at 10,000 x g for 5 min at 4 °C. For lipid-rich samples, carefully avoid/remove the lipid-rich portion and transfer the aqueous phase (supernatant) to a 10kDa MWCO Spin Columns (Cat. # 1997). Centrifuge the sample at 10,000 x g for 10 min at 4 °C and collect the filtrate. For biological fluids: Add 200-500 μl of sample into a 10kDa MWCO Spin Columns (Cat. # 1997). Centrifuge the sample at 10,000 x g for 10 min at 4 °C and collect the filtrate. For all samples: Due to matrix effect in biological samples, an internal standard (spiking) is needed for each sample. For each test sample, prepare 3 parallel sample wells. Add 2-50 μl of samples (2-10 μl of brain tissue sample and 5-25 μl of CSF) into 3 wells in a 96-well clear plate. Label each well as "Sample", "Sample background", "Spike". Dilute D-Amino acid standard to 100 μM by adding 10 μl of the 10 mM stock solution into 990 μl of dH<sub>2</sub>O. Add 2-5 μl of the 0.1 mM D-Amino acid standard into the "spike" wells. Bring the volume of all the wells to 50 μl with DAA Assay buffer. For unknown samples, prepare parallel wells with different dilutions.





- 2. Standard Curve Preparation: Prepare a 100 μM solution of D-Amino acid standard by adding 10 μl of the 10 mM D-Amino acid standard stock to 990 μl of dH<sub>2</sub>O. Add 0, 2, 4, 6, 8, 10 μl of the 100 μM working D-Amino acid standard into a series of wells, generating 0, 200, 400, 600, 800, 1000 pmol of D-Amino acid/well. Adjust the volume to 50 μl/well with DAA Assay buffer
- **3. Reaction Mix:** Mix enough reagents for the number of assays to be performed. Prepare a 5-fold dilution of DAA Probe (e.g. Mix 5 μl of DAA Probe with 20 μl DAA Assay Buffer). For each well, prepare 50 μl Mix containing:

	Reaction Mix	Background Mix
DAA Assay Buffer	42 µl	44 µl
DAA Cofactor	2 µl	2 µl
DAA Enzyme Mix	2 µl	
DAA Developer Mix	2 µl	2 µl
Diluted DAAO Probe	2 µl	2 µl

Mix and add 50  $\mu$ I of the Reaction Mix to each well containing the Standard, Sample and Spike wells. Add 50  $\mu$ I of Background mix to the Sample background wells.

- 4. Measurement: Incubate at 37°C for 2 hours. Protect from light. Measure fluorescence (Ex/Em = 535/587 nm) in a microplate reader.
- 5. Calculation: Subtract 0 Standard reading from all standard readings. Plot the D-Amino acid Standard Curve. Subtract Sample background reading from Sample reading (F<sub>s</sub> = RFU<sub>sb</sub> RFU<sub>sbc</sub>) and subtract Sample background reading from Spike reading (F<sub>spike</sub> = RFU<sub>sbc</sub> RFU<sub>sbc</sub>).

Amount of D-Amino acids in sample wells (B) =  $\frac{F_s}{F_{SDIRe}-F_s}$  ×D-Amino acid Spike (in pmol)

For biological fluids: Sample D-Amino acid Concentration =  $\frac{B}{V} \times D = \text{pmol}/\mu I = \mu M$ 

Where: V is the volume of sample added to the well (in µl)

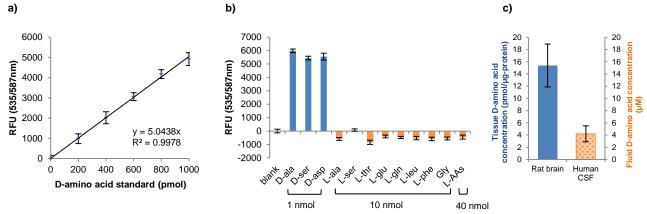
**D** is the sample dilution factor (if applicable, D=1 for undiluted samples)

For tissue samples: Sample D-Amino acid Concentration =  $\frac{B}{V} \times D \times P = \text{pmol/} \mu g$ 

Where: V is the volume of sample added to the well (in µl)

**D** is the sample dilution factor (if applicable, D=1 for undiluted samples)

 ${f P}$  is the sample protein concentration in the untreated samples (µg-protein/µI)



**Figure:** (a) D-Amino acid standard curve; (b) Specificity of the assay: L-AAs were tested at a 10-fold molar excess (each AA: 10 nmol) and 40-fold excess of a mixture of L-AAs (Total: 40 nmol) vs D-AAs (each D-AA: 1 nmol). (c) Estimations of total D-Amino acids in rat brain sample and pooled normal human CSF (15 μl). D-Amino acid concentrations were 15.4 pmol/μg-protein in rat brain and 4.2 μM in Human CSF. Assays were performed following the kit protocol.

## **VIII. RELATED PRODUCTS:**

DL-serine Assay Kit (Fluorometric) (K545)
Pimavanserin Tartrate (B1915)
FAD Colorimetric/Fluorometric Assay Kit (7645)
Ceruloplasmin Activity Colorimetric Assay Kit (K669)
Mutant Alanine Racemase Y354N, Active, Recombinant (P116)
Dextromethrophan (hydrobromide hydrate) (2912)

D-Amino acid oxidase, human recombinant (P1052) Glyoxalase I Activity Assay Kit (Colorimetric) (K591) BRD1 bromodomain (His-Tag), human recombinant (7405) Phospho (Tyr1472) NMDA NR2B Antibody (3616) Anti-Alanine Racemase Antibody (A1380) D (+)-Cycloserine (B1188)

FOR RESEARCH USE ONLY! Not to be used on humans.