

- c. For samples having high protein content, we recommend deproteinizing the samples (tissue or cell lysate or biological fluids) using 10 kDa Spin Column (Cat. # 1997 or equivalent). Add sample to the spin column, centrifuge at 10,000 X g, 4°C for 10 min. Collect the filtrate.
- d. To ensure accurate determination of Glycine in the test samples or for samples having low concentrations of GLY, we recommend spiking samples with a known amount of Glycine Standard (e.g. 0.3 nmol)
- 2. Standard Curve Preparation:** Prepare 1 mM Glycine Standard by adding 10 µl of 100 mM GLY Standard to 990 µl of ddH₂O. Further dilute to 50 µM by adding 50 µl of 1 mM Glycine Standard to 950 µl ddH₂O. Add 0, 2, 4, 6, 8, and 10 µl of 50 µM Glycine Standard into a series of wells in a 96-well plate to generate 0, 0.1, 0.2, 0.3, 0.4 and 0.5 nmol of Glycine/well. Adjust the volume to 50 µl/well with GLY Assay Buffer.
- 3. Reaction Mix:** Dilute GLY Enzyme Mix 10-fold (i.e. 2 µl Gly Enzyme Mix + 18 µl GLY Assay Buffer). Mix enough reagents for the total number of wells to be assayed. For each well, prepare 50 µl of Reaction Mix containing:

	<u>Reaction Mix</u>	<u>*Background Control Mix</u>
GLY Assay Buffer	42 µl	47 µl
Diluted GLY Enzyme Mix	5 µl	----
GLY Developer	2 µl	2 µl
GLY Probe	1 µl	1 µl

Mix well. Add 50 µl of Reaction Mix into Standard and sample wells. Mix.

* For samples having background, add Background Control Mix to background control well(s) and mix.

- 4. Measurement:** Incubate plate at 25°C for 60 min., protected from light. Measure fluorescence (Ex/Em = 535/587 nm) in end point mode.
- 5. Calculation:** Subtract 0 Gly Standard reading from all readings. Plot the Gly Standard Curve. If sample background control is significant, then subtract sample background control reading from sample reading. Apply corrected RFU to Standard Curve to get B nmol Glycine in the sample well.

$$\text{Sample Glycine Concentration (C)} = \text{B/V} \times \text{D nmol/}\mu\text{l or mM}$$

Where: **B** is amount of Glycine in the sample well from Standard Curve (nmol)

V is sample volume added into the reaction well (µl)

D is sample dilution factor

Note: For spiked samples, correct for any sample interference by using the following equation:

$$\text{Glycine amount in spike sample well (B)} = \frac{\text{RFU}_{\text{sample (corrected)}}}{(\text{RFU}_{\text{sample+Gly Std (corrected)}}) - (\text{RFU}_{\text{sample (corrected)}})} * \text{Gly spike (nmol)}$$

Glycine molecular weight: 75 g/mol
1 mM ≡ 1000 µM

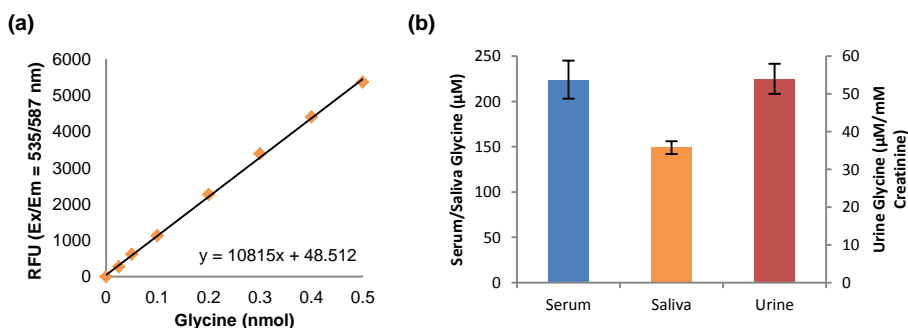


Figure: (a) Glycine Standard Curve. **(b)** Estimation of Glycine concentration in human serum, saliva, and urine. Samples were deproteinized using 10 kDa spin column and diluted using GLY Assay Buffer (Serum: 64-fold; Saliva: 32-fold; Urine: 128-fold). 25 µl of each diluted sample was spiked with 0.3 nmol of Glycine Standard and assayed following the kit protocol. Glycine concentrations are: Serum: 224 ± 21 µM, Saliva: 149 ± 7 µM, Urine: 54 ± 4 µM/mM Creatinine.

VIII. Related Products:

Albumin (Albuminuria) Assay Kit (K550)	Creatinine Colorimetric/Fluorometric Assay Kit (K625)
Albumin Colorimetric (BCG) Assay Kit (K554)	BCA Protein Assay Kit II (K813)
Glutamine Colorimetric Assay Kit (K556)	Glutamate Colorimetric Assay Kit (K629)
Alanine Colorimetric/Fluorometric Assay Kit (K652)	Aspartate Colorimetric/Fluorometric Assay Kit (K552)
Phenylalanine Fluorometric Assay Kit (K572)	Tyrosine Colorimetric Assay Kit (K573)
Hydroxyproline Colorimetric Assay Kit (K555)	Sarcosine Colorimetric/Fluorometric Assay Kit (K636)

FOR RESEARCH USE ONLY! Not to be used on humans.