



Oxalate (Oxalic Acid) Colorimetric Assay Kit

02/14

(Catalog # K663-100; 100 assays; Store at -20°C)

I. Introduction:

Oxalate ($C_2O_4^{2-}$), in the form of Oxalic acid is present in many foods and beverages (e.g. spinach, tea etc.). It accumulates in many plant tissues and play role in regulating pH, osmosis and calcium storage. In animals, oxalate is either absorbed from dietary intake or produced from glycolate metabolism in liver. Under normal conditions, the daily oxalate load can be excreted by kidney. However, hereditary defects can cause an increased level of oxalate, which leads to hyperoxaluria, and results in the formation of kidney stones. Therefore, measurement of oxalate level is useful for the prevention, diagnosis and monitoring of kidney stones. BioVision's Oxalate Assay kit is an easy-to-use, sensitive and high throughput adaptable kit. In this assay, Oxalate reacts with Oxalate Converter & Oxalate Enzyme Mix to form an intermediate, which in turn reacts with a highly specific probe to generate color at 450 nm. The assay kit can detect Oxalate levels lower than 20 μ M.



II. Application:

- Measurement of Oxalate level in various samples.
- Analysis of kidney and liver functions.

III. Sample Type:

- Animal samples: Urine, serum, plasma, kidney, liver etc.
- Plant samples: Vegetable leaves and fruits.

IV. Kit Contents:

Components	K663-100	Cap Code	Part Number
Oxalate Assay Buffer	25 ml	WM	K663-100-1
Oxalate Development Buffer	15 ml	NM	K663-100-2
Oxalate Converter	0.2 ml	Purple	K663-100-3
Oxalate Enzyme Mix (lyophilized)	1 vial	Green	K663-100-4
Oxalate Probe (lyophilized)	1 vial	Red	K663-100-5
Oxalate Standard (lyophilized)	1 vial	Yellow	K663-100-6

V. User Supplied Reagents and Equipments:

- 96-well clear plate with flat bottom
- Multi-well spectrophotometer (ELISA reader)
- Activated charcoal (> 80 mesh)
- Mortar and pestle

VI. Storage Conditions and Reagent Preparation:

Store kit at -20°C, protected from light. Read the entire protocol before performing the assay.

- **Oxalate Assay Buffer** and **Oxalate Development Buffer**: Warm buffers to room temperature before use. Store at 4°C or -20°C.
- **Oxalate Converter**: Aliquot and store at -20°C. Avoid repeated freeze-thaw. Use within two months.
- **Oxalate Enzyme Mix**: Reconstitute with 220 μ l dH₂O. Pipette up and down to dissolve completely. Keep on ice while in use. Aliquot and store at -20°C. Avoid repeated freeze/thaw cycles. Use within two months.
- **Oxalate Probe**: Reconstitute with 220 μ l dH₂O. Pipette up and down to dissolve completely. Stable for two months at -20°C.
- **Oxalate Standard**: Reconstitute with 100 μ l dH₂O to generate 100 mM (100 nmol/ μ l) Oxalate Standard solution. Keep on ice while in use. Store at -20°C. Use within two months.

VII. Oxalate Assay Protocol:

- Sample Preparation**: Liquid samples (e.g. serum) can be measured directly. For plant tissues or fruits (~15 mg), grind in pre-cooled mortar with pestle to break the cell wall until a paste is formed from grinding (for the most efficient extraction, quickly freeze under liquid nitrogen and grind in a pre-cooled mortar using a pestle). Add 150 μ l of ice-cold Oxalate Assay Buffer to tissue/fruit paste/powder and homogenize. Incubate the homogenate for 10 min. on ice and centrifuge at 10,000 X g for 5 min. For animal tissue, homogenize the tissue directly and centrifuge at 10,000 X g for 5 min. Collect the supernatant. Add 1-50 μ l of the supernatant into a 96-well plate and bring the volume to 50 μ l with Oxalate Assay Buffer.

Notes:

- For unknown samples, we suggest doing a pilot experiment & testing several doses to ensure the readings are within the Standard Curve range.
- For urine samples, we recommend collecting samples 24 hrs prior to testing. Collect sample (10 ml) in a bottle containing 10 ml of 10 M hydrochloric acid. Centrifuge at 1,000 X g for 10 min. and either use the supernatant immediately for analysis or save at -20°C for future experiments.
- To reduce background from urine samples, we recommend purifying the urine samples further by mixing 1 ml of urine with activated charcoal (25 mg) [not provided] for 5 min. at room temperature. Centrifuge at 10,000 X g for 5 min. and use 10 μ l of the supernatant for the assay.

