



[Cat. # 7096] at room temperature) and keep on ice for 20 min. Spin down at 10,000 X g for 5 min., carefully remove and discard the supernatant, and resuspend the pellet to the original volume with PDH Assay Buffer.

**2. NADH Standard Curve:** Add 0, 2, 4, 6, 8 and 10  $\mu\text{l}$  of 1.25 mM NADH Standard into a series of wells in 96 well plate to generate 0, 2.5, 5.0, 7.5, 10 and 12.5 nmol/well of NADH Standard. Adjust the volume to 50  $\mu\text{l}$ /well with PDH Assay Buffer.

**3. Reaction Mix:** Mix enough reagents for the number of assays to be performed. For each well, prepare 50  $\mu\text{l}$  Mix containing:

	Reaction Mix	*Background Control Mix
PDH Assay Buffer	46 $\mu\text{l}$	48 $\mu\text{l}$
PDH Developer	2 $\mu\text{l}$	2 $\mu\text{l}$
PDH Substrate	2 $\mu\text{l}$	---

Mix and add 50  $\mu\text{l}$  of the Reaction Mix to each well containing the Standard, Positive Control and test samples.

\* For background correction, add 50  $\mu\text{l}$  of Background Control Mix (without substrate) to sample background control well(s) and mix well.

**4. Measurement:** Measure absorbance immediately at 450 nm in kinetic mode for 10-60 min. at 37°C.

**Note:** Incubation time depends on the pyruvate dehydrogenase activity in samples. We recommend measuring the OD in kinetic mode, and choosing two time points ( $T_1$  &  $T_2$ ) in the linear range to calculate the pyruvate dehydrogenase activity of the samples. The NADH Standard Curve can be read in Endpoint mode (i.e., at the end of the incubation time).

**5. Calculation:** Subtract 0 Standard reading from all readings. Plot the NADH Standard Curve. If sample background control reading is significant, subtract the background control reading from its paired sample reading. Calculate the pyruvate dehydrogenase activity of the test sample:  $\Delta\text{OD} = A_2 - A_1$ . Apply the  $\Delta\text{OD}$  to the NADH Standard Curve to get B nmol of NADH generated during the reaction time ( $\Delta T = T_2 - T_1$ ).

$$\text{Sample Pyruvate Dehydrogenase Activity} = \frac{B}{(\Delta T \times V)} \times D = \text{nmol/min/ml} = \text{mU/ml}$$

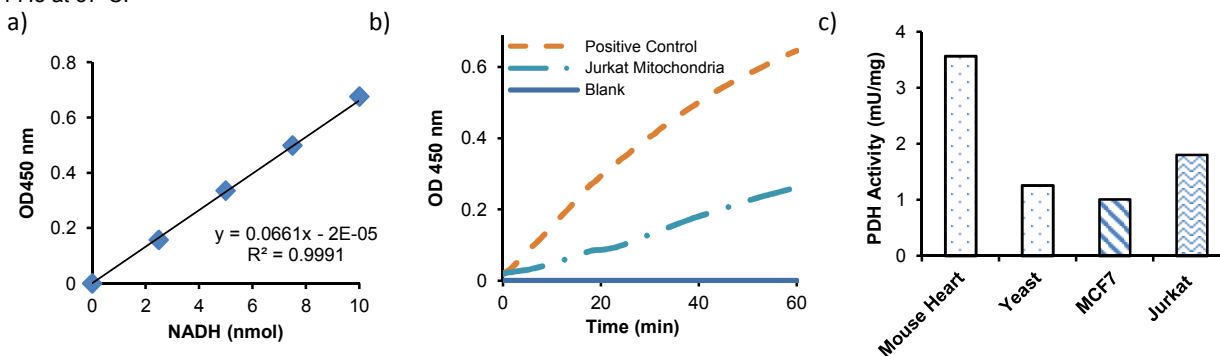
Where: **B** = NADH amount from Standard Curve (nmol).

$\Delta T$  = reaction time (min.).

**V** = sample volume added into the reaction well (ml).

**D** = Dilution Factor

**Unit Definition:** One unit of pyruvate dehydrogenase is the amount of enzyme that generates 1.0  $\mu\text{mol}$  of NADH per min. at pH 7.5 at 37°C.



**Figure:** (a) NADH standard curve; (b) Pyruvate Dehydrogenase activity in Jurkat mitochondria (100  $\mu\text{g}$ ); (c) Pyruvate Dehydrogenase specific activity was calculated in mitochondria prepared from mouse heart (10  $\mu\text{g}$ ), *S. cerevisiae* (10  $\mu\text{g}$ ), MCF-7 cells (30  $\mu\text{g}$ ) or Jurkat cells (35  $\mu\text{g}$ ). Assays were performed following the kit protocol.

#### IX. RELATED PRODUCTS:

Malate Colorimetric Assay Kit (K637)

Pyruvate Colorimetric /Fluorometric Assay Kit (K609)

Citrate Colorimetric/Fluorometric Assay Kit (K655)

Citrate Synthase Activity Colorimetric Assay Kit (K318)

Succinate (Succinic Acid) Colorimetric Assay Kit (K649)

$\alpha$ -Ketoglutarate Colorimetric Assay Kit (K677)

Fumarate Colorimetric Assay Kit (K633)

PicoProbe™ Acetyl-CoA Fluorometric Assay Kit (K317)

Oxaloacetate Colorimetric/Fluorometric Assay kit (K659)

Isocitrate Colorimetric Assay Kit (K656)

Isocitrate Dehydrogenase Activity Assay Kit (K756)

Aconitase Activity Colorimetric Assay Kit (K716)

**FOR RESEARCH USE ONLY! Not to be used on humans.**