



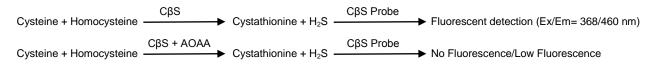
# Cystathionine β Synthase Inhibitor Screening Kit (Fluorometric)

rev 06/19

(Catalog # K695-100; 100 assays; Store at -20°C)

# I. Introduction:

Cystathionine  $\beta$  Synthase (EC 4.2.1.22, C $\beta$ S) is a Pyridoxal 5-Phosphate-dependent enzyme that uses cysteine and homocysteine as substrates to catalyze the formation of H<sub>2</sub>S and cystathionine. C $\beta$ S is well-known for its role in human sulfur metabolism. The overexpression of C $\beta$ S has been implicated in Down Syndrome, and also results in homocystinuria. Homocystinuria leads to a multi-systemic disorder of the connective tissue, muscles, and central nervous system. For that reason, pharmacological inhibitors of human C $\beta$ S present attractive therapeutic potential in order to restore baseline C $\beta$ S activity and blood homocysteine levels. It has been shown that aminooxyacetic acid (AOAA) is an irreversible inhibitor that targets the PLP-binding site of C $\beta$ S. That interaction inhibits the transaminase reaction that would yield cystathionine and H<sub>2</sub>S in wildtype conditions. BioVision's Cystathionine  $\beta$  Synthase Inhibitor Screening Kit is the first available kit that enables customers to screen inhibitors of C $\beta$ S and quantify their therapeutic potential. The kit has a simple, easy-to-follow protocol and is high-throughput adaptable. In this kit, the product hydrogen sulfide reacts with the non-fluorescent azido-functional group to yield a fluorescent amino group (Ex/Em = 368/460 nm). In the presence of AOAA, C $\beta$ S activity is inhibited, which subsequently abolishes the production of H<sub>2</sub>S and thus the fluorescent signal is reduced.



## II. Applications:

• Screening and characterization of inhibitors of Cystathionine β Synthase

### III. Kit Contents:

Components	K695-100	Cap Code	Part Number
CβS Assay Buffer	25 ml	WM	K695-100-1
CβS Probe in DMSO	0.5 ml	Purple	K695-100-2
CβS Substrate	4.0 ml	NM	K695-100-3
Cofactor 1	0.5 ml	Amber	K695-100-4
Cofactor 2	0.5 ml	Orange	K695-100-5
Reducing Agent	1 vial	Yellow	K695-100-6
CβS	0.5 ml	Green	K695-100-7
CβS Inhibitor	100 µl	Clear	K695-100-8

## IV. User Supplied Reagents and Equipment:

- Multi-well fluorescence microplate reader
- 96-well white microtiter plates with flat bottom

### V. Storage Conditions and Reagent Preparation:

Store kit at -20°C, protected from light. Briefly centrifuge vials prior to opening. Read entire protocol before performing the assay.

- CβS Assay Buffer: Equilibrate to room temperature (RT) before running the assay. Store at 4°C.
- CβS Probe in DMSO and CβS Substrate: Light sensitive. Aliquot and store at -20°C. Allow reagents to equilibrate to RT before use.
- Cofactor 1 and Cofactor 2: Aliquot and store at -20°C, stable for at least 4 freeze-thaw cycles.
- Reducing Agent: Reconstitute with 250 μl CβS Assay Buffer. Store at 4°C. Keep on ice during use. Stable for 4 freeze/thaw cycles.
- CβS: Aliquot and store at -20°C. Keep on ice while in use. Use within two months. For long-term use, store CβS in -80°C
- CβS Inhibitor (AOAA, 100 mM in DMSO): Aliquot and store at -20°C. Avoid repeated freeze-thaw. Use within six months.

## VI. C<sub>β</sub>S Inhibitor Screening Protocol:

 Screen Compounds (Co), Inhibitor Control (IC), and Enzyme Control Preparation (EC): Dissolve screen compound into a proper solvent at 100X the desired working concentration. Dilute screen compound (Co) to 20X desired test concentration with CβS Assay Buffer. Add 10 µl each screen compound into a designated well of a white 96-well microplate.

For IC, dilute the C $\beta$ S Inhibitor, AOAA to 10 mM working solution by adding 10  $\mu$ I C $\beta$ S Inhibitor to 90  $\mu$ I C $\beta$ S Assay Buffer. Add 10  $\mu$ I of C $\beta$ S Inhibitor into desired well(s).

For EC, add 10 μl CβS Assay Buffer into well designated as Enzyme Control (EC) (no inhibitor).

**Note:** Solvents (e.g. DMSO) used to solubilize the inhibitors may affect the enzymatic activity. If that is a concern, prepare a solvent control well (SC) with the same final concentration of the solvent(s) as in the Co or IC sample(s).

**2.** Cystathionine  $\beta$  Synthase Enzyme Solution Preparation: For each well, prepare 50 µl of C $\beta$ S enzyme solution.

45 μl CβS Assay Buffer 5 μl CβS

Mix well and add 50 μl/well of CβS Enzyme Solution to each well (with Co, IC, EC, or SC) of the 96-well white microtiter plate. Include sufficient number of wells to test each Co, IC, EC, and SC. Incubate at 30°C for 30 min.





Note: If the plate reader allows, include a 10 sec pulse at the start of the incubation to ensure thorough mixing of enzyme with Co, IC, EC or SC. Also include a 3 sec pulse every 1-5 min.

3. Reaction Mix: During incubation, prepare Reaction Mix. Add Reaction Mix to each well immediately after incubation. Prepare Working Reducing Agent before use. Add 17 µl of Reducing Agent to 483 µl CβS Assay Buffer. Prepare sufficient volume of Reaction Mix for the number of assays to be performed. Prepare 140 µl Reaction Mix per well containing:

	Reaction Mix
CβS Assay Buffer	85 µl
CβS Probe (in DMSO)	2 µl
CβS Substrate	40 µl
Cofactor 1	2 µl
Cofactor 2	1 µl
Working Reducing Agent	10 µl

Add 140 µl Reaction Mix to each well containing Co, IC, EC, or SC.

Note: Do not store the Working Reducing Agent. Always prepare fresh dilution prior to the assay.

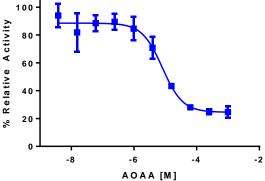
- 4. Measurement: Measure fluorescence immediately at Ex/Em= 368/460 nm in kinetic mode for 40-60 min. at 30°C.
  - **Note:** The enzymatic product ( $H_2S$ ) reacts with the C $\beta$ S probe to yield fluorescence. This may cause a lag phase to appear in the C $\beta$ S Activity Progress Curves.

**5.** Calculation: For each reaction well (Co, IC, EC, SC), choose any two time points ( $t_1$  and  $t_2$ ) in the linear phase of each reaction progress curve. Obtain the corresponding fluorescence values at those points (RFU<sub>1</sub> and RFU<sub>2</sub>). Calculate the slope for all samples, including Enzyme Control (EC), by dividing the net  $\Delta$ RFU (= RFU<sub>2</sub>- RFU<sub>1</sub>) values by the time  $\Delta$ t (=  $t_2$ - $t_1$ ). Calculate % Relative Activity and % Relative Inhibition as follows:

% Relative Activity = 
$$\frac{\text{Slope of SC or EC}}{\text{Slope of EC}} \times 100$$

% Inhibition =  $\frac{\text{Slope of EC-Slope of Co}}{\text{Slope of EC}} \times 100$ 

Note: If the values of the Solvent Control(s) (SC) are significantly different from the Enzyme Control, use the SC values instead of EC values.



**Figure:** (a) Inhibition of C $\beta$ S activity by AOAA Inhibitor Control. IC<sub>50</sub> was determined to be 8.0 ± 0.11  $\mu$ M. Assays were performed following the kit protocol.

## VII. Related Products:

Cysteine Assay Kit (Fluorometric) (K558)Cystathionine β-Synthase A<br/>Cystathionine β Synthase Activity Assay Kit (Fluorometric)Cystathionine β-Synthase A<br/>Cystathionine β-Synthase,<br/>Cystathionine β-Synthase,<br/>CBS Antibody (Center) (67<br/>CBS Antibody (NT) (6729)Adenosylhomocysteinase (AHCY) Activity Fluorometric Assay Kit (807)CBS Antibody (NT) (6729)

Cystathionine  $\beta$ -Synthase Antibody (A1112) Cystathionine  $\beta$ -Synthase, human recombinant (7844) CBS Antibody (Center) (6728) CBS Antibody (NT) (6729)

## FOR RESEARCH USE ONLY! Not to be used on humans.

THE REPORT OF A DECART TO THE MANY OR ADDRESS TO THE MANY OF A DATE OF A DAT