

MMP-3 Inhibitor Screening Kit (Fluorometric)

(Catalog #K793-100; 100 assays; Store kit at -20°C)

I. Introduction:

The matrix metalloproteinase-3 (MMP-3, stromelysin-1) exhibits a number of activities that would make it a particularly good tumor promoter. Like several other MMPs, MMP-3 was first cloned and later re-cloned as a cancer-specific gene. In addition to degrading numerous extracellular matrix components, MMP-3 can activate gelatinase B, the collagenases and several serpin-type serine proteinase inhibitors. Moreover, it can release a number of cell surface molecules, including E-cadherin, a known contributor to cancer development. In BioVision's MMP-3 Inhibitor Screening Assay Kit, MMP-3 hydrolyzes a specific FRET substrate to release the quenched fluorescent group Mca, which can be detected fluorometrically at Ex/Em = 325/393 nm. The kit provides a rapid, simple, sensitive, and reliable test suitable as a high throughput screening assay of MMP-3 inhibition. In addition, we also offer a human recombinant MMP-3 enzyme (Biovision #7783) and a MMP-3 Activity Assay Kit (Biovision #K783-100), separately.

II. Kit Contents:

Components	100 assays	Cap Code	Part Number
MMP-3 Assay Buffer	25 ml	WM	K793-100-1
MMP-3 Substrate	200 µl	Red	K793-100-2
MMP-3 Enzyme (lyophilized)	1 vial	Green	K793-100-3
Inhibitor Control (0.1 mM GM6001)	20 µl	Purple	K793-100-4

III. Storage and Handling:

Store the kit at -20°C, protect from light. Allow Assay Buffer to warm to room temperature before use. Briefly centrifuge vials before opening. Read the entire protocol before performing the assay.

IV. Reagent preparation:

MMP-3 Enzyme: Reconstitute the MMP-3 enzyme into 220 µl assay buffer. Aliquot and store the MMP-3 stock solution at -80°C. Avoid repeated freeze/thaw cycles. Use within one week.

V. MMP-3 Inhibitor Screening Assay Protocol:

- Inhibitor Compounds, Inhibitor Control and Blank Control Preparations:** Dissolve candidate compounds into proper solvent. Dilute to 2X concentration with Assay Buffer. Add 50 µl diluted compounds solution into MMP-3 enzyme wells. For Inhibitor Control, use 2 µl and dilute to 50 µl with Assay Buffer. Use Assay Buffer alone for Blank Control. Mix well.
- MMP-3 Enzyme Solution:** Mix enough reagents for the number of assays to be performed. For each well, prepare a total 50 µl MMP-3 enzyme solution.

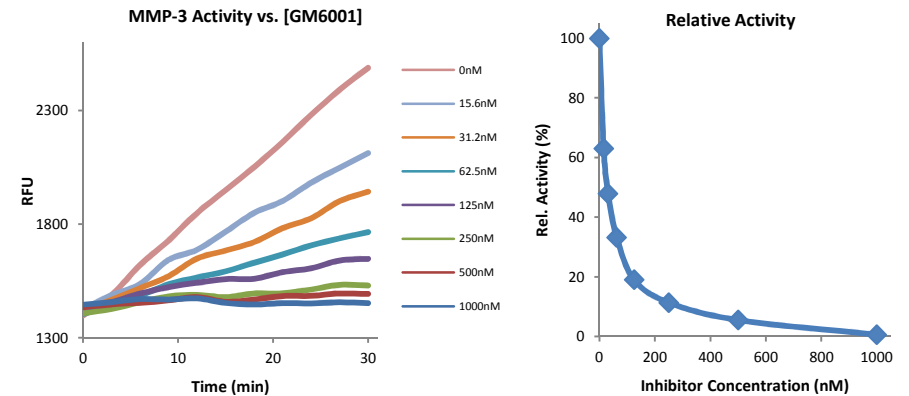
48 µl Assay Buffer
2 µl MMP-3 Stock Solution

Add 50 µl of the MMP-3 enzyme solution to each well. Incubate candidate compounds-enzyme mixes, inhibitor control-enzyme mix and blank control for 10 min at 37°C.

- Substrate:** Dilute Substrate 1:5 with Assay Buffer. Add 10 µl diluted substrate into each well. Mix well.
- Measurement:** Read Ex/Em = 325/393 nm R₁ at T₁. Read R₂ again at T₂ after incubating the reaction at 37°C for 30 min, protect from light. The RFU of fluorescence generated by hydrolyzation of substrate is ΔRFU = R₂ - R₁.
- Calculation:** Set the ΔRFU of blank control as the 100 %, and calculate the relative activity remaining with candidate compounds as follows.

$$\text{Activity Remaining} = \frac{\Delta\text{RFU of candidate}}{\Delta\text{RFU of blank}} \times 100 \%$$

It is recommended to read kinetically to choose the R₁ and R₂ within a linear range.



RELATED PRODUCTS:

- Human recombinant proteins: MMP-1, -2, -3, -8, -9, -11, -12, -13
- MMP antibodies to: MMP-1, -2, -3, -8, -9, -11, -12, -13, -17, -19
- MMP blocking peptides to: MMP-3, -8, -9, -11, -12
- MMP-3 Inhibitor GM6001
- MMP-3 Activity Assay Kit

FOR RESEARCH USE ONLY! Not to be used on humans.

GENERAL TROUBLESHOOTING GUIDE:

Problems	Cause	Solution
Assay not working	<ul style="list-style-type: none"> • Use of ice-cold assay buffer • Omission of a step in the protocol • Plate read at incorrect wavelength • Use of a different 96-well plate 	<ul style="list-style-type: none"> • Assay buffer must be at room temperature • Refer and follow the data sheet precisely • Check the wavelength in the data sheet and the filter settings of the instrument • Fluorescence: Black plates (clear bottoms) ; Luminescence: White plates ; Colorimeters: Clear plates
Samples with erratic readings	<ul style="list-style-type: none"> • Use of an incompatible sample type • Samples prepared in a different buffer • Cell/ tissue samples were not completely homogenized • Samples used after multiple free-thaw cycles • Presence of interfering substance in the sample • Use of old or inappropriately stored samples 	<ul style="list-style-type: none"> • Refer data sheet for details about incompatible samples • Use the assay buffer provided in the kit or refer data sheet for instructions • Use Dounce homogenizer (increase the number of strokes); observe for lysis under microscope • Aliquot and freeze samples if needed to use multiple times • Troubleshoot if needed • Use fresh samples or store at correct temperatures until use
Lower/ Higher readings in Samples and Standards	<ul style="list-style-type: none"> • Improperly thawed components • Use of expired kit or improperly stored reagents • Allowing the reagents to sit for extended times on ice • Incorrect incubation times or temperatures • Incorrect volumes used 	<ul style="list-style-type: none"> • Thaw all components completely and mix gently before use • Always check the expiry date and store the components appropriately • Always thaw and prepare fresh reaction mix before use • Refer datasheet & verify correct incubation times and temperatures • Use calibrated pipettes and aliquot correctly
Readings do not follow a linear pattern for Standard curve	<ul style="list-style-type: none"> • Use of partially thawed components • Pipetting errors in the standard • Pipetting errors in the reaction mix • Air bubbles formed in well • Standard stock is at an incorrect concentration • Calculation errors • Substituting reagents from older kits/ lots 	<ul style="list-style-type: none"> • Thaw and resuspend all components before preparing the reaction mix • Avoid pipetting small volumes • Prepare a master reaction mix whenever possible • Pipette gently against the wall of the tubes • Always refer the dilutions in the data sheet • Recheck calculations after referring the data sheet • Use fresh components from the same kit
Unanticipated results	<ul style="list-style-type: none"> • Measured at incorrect wavelength • Samples contain interfering substances • Use of incompatible sample type • Sample readings above/below the linear range 	<ul style="list-style-type: none"> • Check the equipment and the filter setting • Troubleshoot if it interferes with the kit • Refer data sheet to check if sample is compatible with the kit or optimization is needed • Concentrate/ Dilute sample so as to be in the linear range
<p>Note: The most probable list of causes is under each problem section. Causes/ Solutions may overlap with other problems.</p>		