



little effect on IDO1 activity at a final concentration of  $\leq 0.5\%$ ). For higher concentrations or solvents other than DMSO, we recommend preparing a solvent control (SC) well with the same final concentration of solvent used to solubilize TCs and using this well to define 100% activity if different from no inhibitor control well(s).

## 2. Assay Reaction Preparation:

a. Prepare a 2X Reaction Premix by diluting the 100X Antioxidant Mix in IDO1 Assay Buffer at a 1:50 ratio. Make a sufficient amount of 2X Reaction Premix to add 50  $\mu\text{l}$  to each reaction well (for example, for 10 reactions, mix 10  $\mu\text{l}$  of the 100X Antioxidant Mix with 490  $\mu\text{l}$  IDO1 Assay Buffer).

**Note:** Remember to account for any control reactions (such as background control, no inhibitor/solvent control and positive inhibition control wells) when calculating the amount of 2X Reaction Premix to prepare.

b. Set up the assay reactions according to the table below. Prepare reaction wells containing test compounds, as well as corresponding no inhibitor control (which may also serve as a solvent control (SC), if desired) and background control wells. A positive inhibition control well may also be prepared using the IDO1 Inhibitor (IDO5L). Dilute the stock at a 1:100 ratio by adding 10  $\mu\text{l}$  of the reconstituted 1 mM solution to 990  $\mu\text{l}$  IDO1 Assay Buffer, yielding a 10  $\mu\text{M}$  working solution (10X final concentration) and add 10  $\mu\text{l}$  of the 10X solution to each positive inhibition control well. Adjust the volume of all TC and control wells to 90  $\mu\text{l}$  with IDO1 Assay Buffer.

|                            | No Inhibitor/SC  | +Test Compound   | Background Control | Positive Inhibition Control |
|----------------------------|------------------|------------------|--------------------|-----------------------------|
| Reaction Premix (2X)       | 50 $\mu\text{l}$ | 50 $\mu\text{l}$ | 50 $\mu\text{l}$   | 50 $\mu\text{l}$            |
| Test Compound (10X)        | —                | 10 $\mu\text{l}$ | —                  | —                           |
| Recombinant Human IDO1     | 10 $\mu\text{l}$ | 10 $\mu\text{l}$ | —                  | 10 $\mu\text{l}$            |
| IDO1 Inhibitor IDO5L (10X) | —                | —                | —                  | 10 $\mu\text{l}$            |
| IDO1 Assay Buffer          | 30 $\mu\text{l}$ | 20 $\mu\text{l}$ | 40 $\mu\text{l}$   | 20 $\mu\text{l}$            |

**Note:** For solvent control (SC), use 30  $\mu\text{l}$  IDO1 Assay Buffer containing the appropriate solvent at 3.33X final concentration.

c. Pre-incubate the plate for 10 min at room temperature to allow test compounds to interact with IDO1. The pre-incubation time can be optimized for other test compounds depending on mechanism of action. During the pre-incubation, prepare IDO1 Substrate solution by adding 100  $\mu\text{l}$  of the reconstituted 10 mM L-tryptophan solution to 900  $\mu\text{l}$  IDO1 Assay Buffer, generating a 1 mM solution (10X final concentration). Add 10  $\mu\text{l}$  of the 1 mM solution to each assay well, for a final reaction volume of 100  $\mu\text{l}$ /well. Incubate the plate at 37°C in a dark environment for 45 min (we recommend incubating with gentle shaking to ensure adequate mixing of well contents).

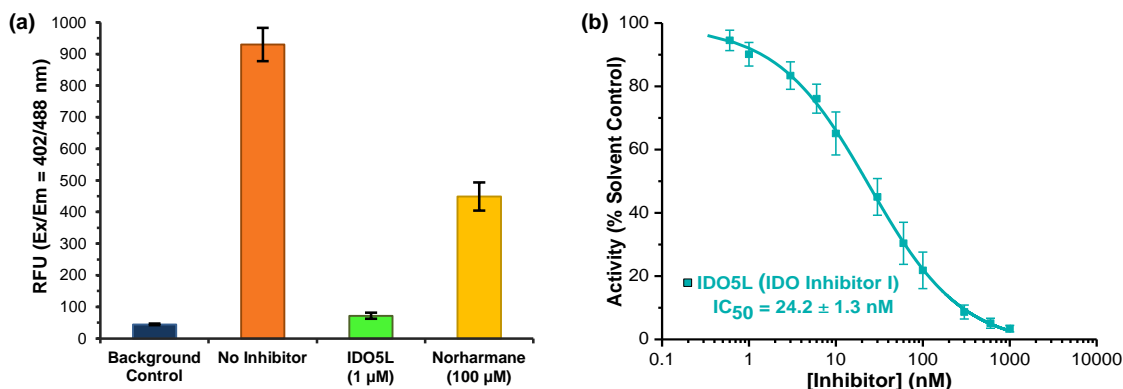
3. **Measurement:** Add 50  $\mu\text{l}$  of the Fluorogenic Developer Solution to each well and tightly seal the plate with the sealing film. Incubate the sealed plate at 45°C in a dark environment for 3 hrs, then allow the plate to cool to room temperature for at least 1 hr. Briefly centrifuge the plate before unsealing. Carefully remove the plate sealing film and measure the fluorescence intensity (Ex/Em = 402/488 nm) of all wells in end-point mode.

### Notes:

- The fluorescent signal generated by the Fluorogenic Developer Solution is stable for 8-12 hrs after the incubation at 45°C, as long as the plate remains sealed and protected from light.

4. **Calculation:** For each reaction well, including no inhibitor/solvent control and positive inhibition controls, subtract the fluorescence intensity of the background control well to determine background-corrected fluorescence (denoted by  $F$ ). Calculate percent inhibition versus no inhibitor/solvent control (SC) due to the test compound (TC) or IDO5L positive inhibition control using the following equation:

$$\% \text{ Relative Inhibition} = \frac{F_{SC} - F_{TC}}{F_{SC}} \times 100\%$$



**Figure:** (a) Measurement of IDO1 inhibition in presence and absence of 1  $\mu\text{M}$  of IDO5L (a potent, highly selective competitive IDO1 inhibitor) and 100  $\mu\text{M}$  of norharmane (a natural product that acts as a weak IDO1 inhibitor). The no inhibitor reaction contained assay buffer with 0.1% DMSO (v/v) as a solvent control. (b) Dose-response curve for IDO1 inhibition by the included selective IDO1 Inhibitor IDO5L. The  $IC_{50}$  value ( $24.2 \pm 1.3$  nM) was derived by 4-parameter logistic curve fitting with each point representing the mean  $\pm$  SEM of at least 3 replicates. All assays were performed according to the kit protocol.

## VII. RELATED PRODUCTS:

|   |                                |
|---|--------------------------------|
| Indoximod (2746)                              | IDO1 Activity Assay Kit (K972) |
| INCB24360/Epacadostat (9477)                  | NLG919 (9407)                  |
| Interferon- $\gamma$ Human Recombinant (4116) | Menadione (2747)               |
| IDO1 Inhibitor I (2745)                       |                                |

**FOR RESEARCH USE ONLY! Not to be used on humans.**