# Protein L Magnetic Beads

**CATALOG #:** 6537-1

**AMOUNT:** 1 ml

**PREPARATION:** Protein L Magnetic Beads are prepared by covalently coupling Recombinant Protein L (contains five Ig kappa light chain binding domains, BV catalog # 6530) to 4% cross-linked magnetically beaded agarose. The coupling technique is optimized to give a high binding capacity. The capacity of IgG binding is generally greater than 10 mg of human IgG per ml of wet gel.

**CONTENTS:** Supplied as a 50% slurry in 20 % Ethanol/dW.

**TECHNICAL SPECIFICATIONS:**

| Parameter | Description |
|---|---|
| Support Characteristics | Paramagnetic, spherical, 6 % cross-linked agarose |
| Ligand | Recombinant Protein L |
| Particle Size | 75 – 150 μm |
| Binding Capacity | Generally >10 mg human IgG/ml wet beads |
| Working Temperature | Room temperature |
| Storage Solution | 20 % Ethanol/dW |
| Storage Temperature | 4 – 8 °C |
| Stability | Stable, as supplied, for at least 1 year |

**FEATURES:** Easy to use, high-binding capacity, non-adherent beads. Useful for immunoprecipitation and enrichment of antibodies. High affinity for kappa-light chain containing Ig antibodies from a variety of species. Protein L binds to all IgG subclasses from human, mouse and rat species. It also binds to human, mouse and rat IgM, IgA, IgE and IgD, as well as FAB with Kappa-light chains. Protein L is superior for binding chicken, hamster and pig IgG.

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***FOR RESEARCH USE ONLY! Not to be used on humans.***

**SUGGESTED PROTOCOL:**

Prepare the antibody solution by diluting the required amount of antibody in binding buffer before running the protocol.

1. Magnetic Bead Preparation (perform three times)
   a. Dispense the required amount of magnetic beads into a 1.5 ml microfuge tube.
   b. Place the tube in the magnetic rack and remove the storage solution.
   c. Add 500 µl binding buffer.
   d. Resuspend the beads.
   e. Remove the liquid
2. Antibody Capture
   a. Immediately add the antibody solution.
   b. Resuspend and mix (slow end-over-end) for at least 15 minutes.
   c. Remove the liquid.
3. Washing
   a. Add 500 µl Binding Buffer containing 0.5 M NaCl; Remove the liquid.
   b. Add 500 µl Binding Buffer; Remove the liquid.
4. Target Binding
   a. Add sample diluted in binding buffer.
   b. Incubate with slow end-over-end mixing for up to 60 minutes.
   c. Remove and collect unbound fraction.
5. Washing ( perform three times)
   a. Add 500 µl wash buffer
   b. Remove liquid (save washes to troubleshoot)
6. Elution (perform three times)
   a. Add 2 volumes elution buffer (vs. bead volume).
   b. Completely resuspend beads and incubate at least 2 minutes.
   c. Remove and collect elution fraction.

**RECOMMENDED BUFFER EXAMPLES:**

**Binding buffer:** 50 mM Tris, 150 mM NaCl, pH 7.5

**Wash buffer:** 50 mM Tris, 150 mM NaCl, pH 7.5 (or add 1% Octylglucoside to this buffer)
(Could also try 1X PBS as both binding and wash buffer)

**Elution buffer:** 0.1 M -0.2 M Glycine pH 2.5-3.1 (or 0.1 M citric acid, pH 2.5-3.1 or 2.5 % Acetic Acid)

**RELATED PRODUCTS:**

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|---|---|---|
| Recombinant Protein A | Protein A Sepharose | Protein A Magnetic Beads |
| Recombinant Protein G | Protein G Sepharose | Protein G Magnetic Beads |
| Recombinant Protein L | Protein L Sepharose | Protein L Magnetic Beads |
| Recombinant Protein A/G | Protein A/G Sepharose | Protein A/G Magnetic Beads |
| Recombinant Protein A/G/L | Protein A/G/L Sepharose | Protein A/G/L Magnetic Beads |
| Protein G Polyclonal Antibody | Protein G-Biotin | Protein G-FITC |
| Protein A Polyclonal Antibody | Protein G Coated Plates | |
| Protein L Polyclonal Antibody | | |

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