Hi-Bind™ Cibacron Blue-Agarose Beads (Albumin Removal Beads)

CATALOG #: 7923-10 10 ml
7923-50 50 ml

FORMULATION: Provided as 50% aqueous slurry containing 20% ethanol.

Ligand Density: 8-10 µmol Cibacron Blue 3G-A per ml of drained gel/resin.

BINDING CAPACITY: The binding capacity of the Hi-Bind™ Cibacron Blue 3G-A-Agarose beads is >18 mg of Human Serum Albumin per ml of the drained gel/resin.

DESCRIPTION: Cibacron Blue has been shown to bind dehydrogenases, kinases, serum albumin, interferons, several plasma proteins, and a variety of other proteins. It is a popular ligand for dye-ligand affinity chromatography. BioVision's Cibacron Blue-Agarose beads are prepared by covalent conjugation of Cibacron Blue 3G-A to cross-linked (4%) Agarose beads by a proprietary method. The beads can be used as a purification matrix for a variety of Cibacron Blue dye-binding proteins. In addition, they are useful for enrichment of low abundant non-Cibacron Blue binding proteins.

APPLICATIONS: BioVision's Hi-Bind™ Cibacron Blue-Agarose beads can be used as an affinity purification matrix of variety of proteins for subsequent ELISA, immunoassays and other downstream analyses such as gel electrophoresis, functional assays, etc.

STORAGE CONDITIONS: Store at 4°C. Do not freeze the resin.

GENERAL PROTOCOL: The protocol given below is a general protocol for purifying proteins using Hi-Bind™ Cibacron Blue-Agarose beads. Certain modifications may be necessary to the protocol, depending upon the type of protein.

Suggested Binding Buffer: 50 mM Tris buffer, pH 8.0 or PBS.

Suggested Elution Buffer: Choose an appropriate elution buffer from below:

- 50 mM Tris, 1 M NaCl, pH 8.0.
- Other buffer containing a competing ligand.

FOR RESEARCH USE ONLY! Not to be used in humans.

1. **Column Preparation:** Carefully pack 1-5 ml of resin slurry in a disposable column avoiding air bubbles and allow the buffer to drain through. Wash the resin with 4x5 column volumes (CV) of Binding Buffer. Do not allow the resin to dry. Close the column outlet.
2. **Sample Preparation:** Load sample on the resin (If necessary dilute to 5 ml with binding buffer). Close the column with a cap.
3. Allow the sample to bind to the resin for 30 min by mixing the suspension on a rotary shaker or intermittently by hand.
4. After 30 min, open the column (both top and outlet) and collect the flow-through fraction. Wash the column with 5-10 CV of binding buffer. Combine washes together and concentrate if necessary.
5. Elute the protein from the resin using appropriate elution buffer.
6. Analyze the flow through, wash and eluted protein, by SDS-PAGE, UV or any other functional assay.
7. To regenerate/store column: A, Wash with 5 volumes of Elution Buffer; B, Wash with 5 volumes of distilled water; C, Store column in 20 % Ethanol/H_2O at 4°C.
8. To clean/store column: A, Wash with 5 volumes of 0.2 N NaOH or 6 M Guanidine HCl; B, Wash with 5 volumes of distilled water; C, Store column in 20 % Ethanol/H_2O at 4°C

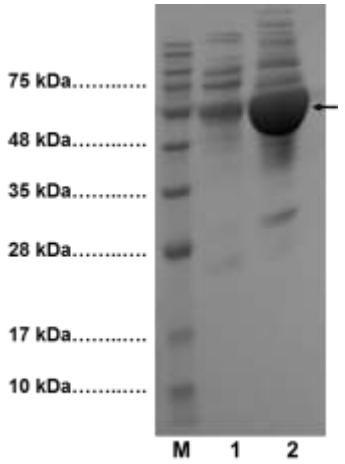

Figure: Hi-Bind™ Cibacron Blue-Agarose beads were used to isolate albumin from plasma sample (15% SDS-PAGE):
M: Protein Marker, 1: Flow-through fraction containing unbound enriched plasma proteins, 2: Eluted fraction (primarily containing albumin, indicated by an arrow) from the beads using BioVision's proprietary buffer.

RELATED PRODUCTS:

- Albumin, Human Plasma **(7546-1)**
- Human Serum Albumin **(4016-1, -10)**
- Hi-Bind Ni QR Agarose Beads **(6562-1, -10, -100, 500)**
- Protein A-Agarose **(6526-1, -5, -25, -100)**
- Heparin Sepharose Column **(6554-1, -5)**

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